



Study the Expression level of Annexin A2 Gene in Oviduct Epithelial Cells of Iraqi Cows with Endometritis

Dhia Hussain J. Al-Delemi*[✉] Mohammed Dhafer T. Alobaidi

Dept.of Surgery and Obstetrics / College of Veterinary Medicine , Al-Qadisiyah University / Iraq*

<https://orcid.org/0000-0002-0874-9679> [✉]

Scopus Author ID: 57216729898

ResearcherID:X-6712-2018

Submitted: March 07, 2025

Revised: March 20, 2025

Accepted: March 23, 2025

Correspondence

Dhia H, J. Al-Delemi

dheyaa.aldulaimi@quedu.iq

Abstract

The present study carried out to study the Annexin A2 gene Expression in oviduct epithelial cells of Iraqi cows with endometritis in comparison with it in the oviduct epithelial cells of healthy cows. Fifty uterine sample were collected from cows with macroscopic change in the uterus, samples were divided into two groups (molecular and histopathology) group . the samples were taken from oviduct ampulla immediately plunged into liquid nitrogen; and stored at -80 °C until the molecular study. The histological examination of 50 endometrial samples of the cows reproductive tract revealed that 30 samples exhibited histological changes associated with endometritis, while the 20 samples were normal endometrium . The molecular study involved RNA extraction from oviduct tissue ampulla and RNA concentration measurement by Quantus™ Fluorometer , cDNA syntheses and Quantitative Reverse transcriptase PCR (RT-qPCR) Preparation, The gene expression analysis of ANNEXIN A2 using RT-qPCR technology revealed a significant up-regulation in cows oviduct epithelial cell ampulla within endometritis compared to oviduct epithelial cells ampulla of normal endometrium cows (p-value< 0.0022) and significantly different (P < 0.01) the expression level (fold change) in oviduct with endometritis cows was (1.969) whereas oviduct with normal endometrium cows (1.076). In conclusion ,the result indicated that presence of gross lesion changes does not be necessarily indicate microscopic pathological change, Annexin A2 up-regulation in oviduct epithelial cells ampulla of cows with endometritis. This study identified potential marker gene for fertility in dairy cattle.

Keywords : Cow, Endometritis, Oviduct, Epithelial Cell, Annexin A2.gene.

©Authors, 2025, College of Veterinary Medicine, University of Al-Qadisiyah. This is an open access article under the CC BY 4.0 license (<http://creativecommons.org/licenses/by/4.0/>).

Introduction

One of the main reasons for infertility and financial losses in the dairy industry is inflammation of uterine endometrium called endometritis, which frequently occurs after calving are diagnosed between 3 and 5 weeks postpartum [1]. There are several type of pathogenic bacteria lead to development cow endometritis including Staphylococcus, Streptococcus, Escherichia, Corynebacterium, Pseudomonas, Proteus, Necrobacillus, Pseudomonas aeruginosa, Campylobacter genitalia, Haemophilus, Bacillus pyogenes, Bacterium burgeri, Trueperella, Fusobacterium, and Prevotella, [2].the classification of cow endometritis to clinical endometritis characterized with purulent vaginal discharge without systemic

illness and subclinical endometritis without any clinical sign [3] .oviduct is small tubular structure linked uterus with ovary and it is important in many reproductive events such as fertilization, oocyte final maturation and early embryo development[4] anatomically contain three regions, isthmus, ampulla and infundibulum [5] oviduct epithelial cells play important role for fertility because it is prolong the life span of sperm, and the sperm binds with the ova in oviduct[6], so any problem in oviduct causes infertility [7] ,the one of main protein in cow oviduct is Annexin A2 gene the function of Annexin A2 gene in cow oviduct linked with binding of sperm in epithelial cell of oviduct and formation functional reservoir[8] its found the Annexin A2 gene have function in



interaction between gamete, early embryo with oviduct epithelial cell is crucial for fertilization and early embryo development [9]. Annexin A2 protein coding gene belongs to the Annexin family; it is a multifunctional calcium²⁺ (Ca²⁺) and phospholipid-binding protein [10], playing an important role in regulating inflammatory response and the infection process of bacteria, viruses, and other pathogens [11]. Annexin A2 plays an integrant role in adhesion of bacteria and in the absence of Annexin A2 there is a decrease of bacterial adhesion [12]. During infection, it facilitates the adhesion and internalization of bacteria and viruses, elevating their replication and release, which is harmful to the host [13]. The present study was carried out to study the Annexin A2 gene expression in oviduct epithelial cells of Iraqi cows with endometritis in comparison with it in the oviduct epithelial cells of healthy Iraqi cows.

Material and Methods

Ethical approved

Ethical approval was granted according to the committee for Ethical Scientific Research at the College of Veterinary Medicine, University of Al-Qadisiyah (Approval number P.G/872 at 24/2/2025).

Sample collection

Collected all the research samples, total 50 uterine samples with uterine macroscopically change, from the female reproductive system of cows slaughtered at the slaughterhouse in al-qadisiyah governorate. The collection was conducted twice a week for five months, from 1/10/2024 to 1/2/2025. The samples were taken from endometrium and oviduct ampulla. The sample was divided into two groups (molecular and histopathology) groups. The histological and molecular study were done in the labs of the Veterinary Medicine College / University of Al-Qadisiyah.

Samples for molecular study

Samples were taken from cows' oviducts, and sealed in a plastic bag, and transported by box containing ice. They were cleaned from blood clots and extra tissues by washing in phosphate buffer saline (PBS). Then, a surgical scalpel was used to extract 3cm³ from the ampulla, and the sample was put in a Cryovial tube to store in a liquid nitrogen tank until further processing (Gene Expression). Isolation of oviduct epithelial cell protocols according to [14].

Samples for histopathology study

The samples were taken from the cow's endometrium, used a surgical scalpel to cut 1cm³ from the uterine endometrium layer, washed in normal saline, and put in a cup filled with 10% formalin buffer. According to [15], the sample preparation for histological sectioning according to [16] and [17] methods, 50 samples (1cm³) were taken from cow

uterine endometrium, sections of endometrial specimens for light microscopy, 5-7 μm thick, were stained with Hematoxylin and Eosin.

RNA extraction and cDNA synthesized

For Annexin A2 and GAPDH genes, a set of primer was synthesized by Macrogen (South Korea) according to [18] table (1). Total RNA extraction according to the manufacturer's protocol using a silica gel column-based spin column method using the kit ADDBIO/Korea kit, RNA concentration measurement by QuantusTM Fluorometer (Promega, USA). cDNA synthesis using the kit from ADDBio (Korea). The relative expression of genes of interest was normalized using GAPDH gene as a reference gene and calculated using the 2-ΔΔCt method Schmittgen and Livak [19]. Quantitative real-time polymerase chain reaction (qPCR) was performed using Real time qPCR machine (Biorad /USA) using AddScript RT-qPCR Syber master (AddBio, Korea).

Statistical analysis

Statistical analyses were performed using the Graph Pad Prism software v10.4.1. Gene expression level between oviduct ampulla of cow with endometritis and healthy endometrium groups were analyzed using unpaired t-test, mean, STD. Differences with $p < 0.01$ were considered statistically significant. According to [14].

Result

The histological examination of 50 endometrial samples from the cows reproductive tract revealed that 30 samples exhibited histological changes associated with endometritis, while the 20 samples were normal endometrium. The histological study of the endometrium in cows revealed several tissue changes confirming the presence of endometritis: the observed histological changes included glandular part of endometrium clear sloughing of epithelium. The epithelial cells show necrotizing process, liquefactive necrosis and severe damage in the endometrium myofiber layer, with infiltration of inflammatory cells (mainly macrophages). Figure (1)

Gene expression ANNEXIN A2

The gene expression analysis of ANNEXIN A2 using RT-qPCR technology revealed a significant up-regulation in cows oviduct epithelial cells ampulla within endometritis compared to oviduct epithelial cells ampulla of normal endometrium cows (p -value < 0.0022) and significantly different ($P < 0.01$) the expression level (fold change) in oviduct with endometritis cows was (1.969) whereas oviduct normal endometrium cows (1.076) with a mean difference (0.8935 ± 0.2764 mean \pm SEM). Figure (2)

Table 1. Real-time PCR primers

Gene	Primer	sequence 5'-3'	References
GAPDH	Forward	ACCCAGAAGACTGTGGATGG	[18]
	Reverse	ACGCCTGCTTCACACCTTC	
Annexin A2	Forward	CGTGCTCCAGCTAACAGTTCT	[18]
	Reverse	GGAAAGCCAGGTAATGCCTA	

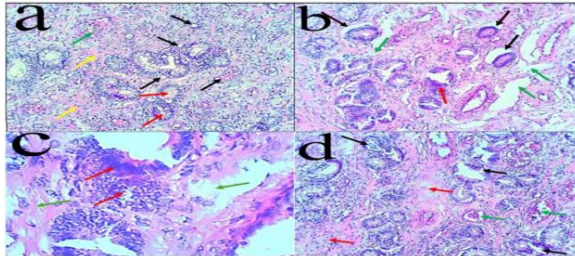


Figure 1. histological section of cow endometrium .a The glandular part of endometrium shows vacuolations in the epithelial cells (Black arrows) with narrowing of glands lumen (Red arrows). The section shows clear blood vessels congestion (Green arrows). The section shows severe damage in the endometrium myofiber layer (Yellow arrows).b The glandular part of endometrium shows clear sloughing of epithelium from glandular basement membrane (Black arrows). The epithelial cells show necrotizing process (liquefactive necrosis, Red arrow). The section shows severe damage in the endometrium myofiber layer (Green arrows). C The epithelial cells show necrotizing process (liquefactive necrosis, Red arrows). The section shows severe damage in the endometrium myofiber layer (Green arrows).d The glandular part of endometrium shows necrotizing lesion (Black arrows) in the epithelial cells and myofibers layer (Red arrows) with infiltration of inflammatory cells (mainly macrophages). The section shows clear blood vessels congestion (Green arrows).

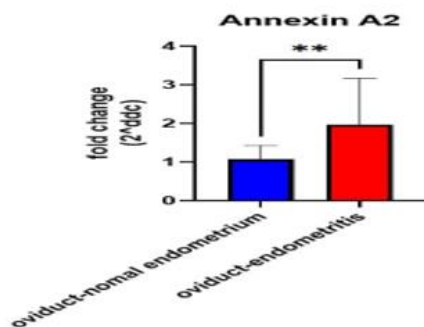


Figure 2. showed up-regulation of Annexin A2 gene expression in oviduct epithelial cell ampulla with endometritis compared with oviduct epithelial cell ampulla of normal endometrium cows (p-value <0.0022)

Discussion

The histological examination of 50 endometrial samples from the cows reproductive tract revealed that 30 samples exhibited histological changes associated with endometritis, while the 20 samples were normal endometrium. The histological study showed the presence of severe damage in the endometrium myofiber layer ,the glandular part of endometrium shows clear sloughing of epithelium necrotizing process liquefactive necrosis , The glandular part of endometrium shows vacuolations in the epithelial cells blood vessels congestion infiltration of inflammatory cells mainly macrophages this result agreed with [20] which show that necrobiosis of the epithelial layer of the mucosa, cellular infiltration with shaped elements of blood in the functional layer, swelling of the cells of the uterine gland edema of the stroma of the functional layer of the endometrial mucosa, moreover, the presence of gross lesions does not necessarily indicate the presence of Histopathological changes ,this observation agreed with the study of [21]in female camels which mention that the gross examination of 247 samples from female reproductive tract revealed that only 67 exhibited macroscopic pathological change, and among them only 25 samples tested positive.

The results of the gross and histological lesion examination of samples reveal that there is several factors may effect on the gross and histological lesion examination like truma, light during slaughter, animal injuries, or increased blood flow , congestion due to vascular pressure and physiological state of female reproductive system .

The present study using RT-qPCR technology showed up-regulation in gene expression in oviduct epithelial cell ampulla of cows with endometritis fold change 1.969 compared with oviduct epithelial cell ampulla cows with normal endometrium fold change 1.076 with a mean difference (0.8935 ± 0.2764 mean ± SEM) p value <0.0022, this result agree with the results of [22]which study Annexin A2 in cow endometrium which show that there is strong positive correlation with bacterial infection of endometrium and found there is up-regulation in cow with endometritis in his studies by using 2D gel electrophoresis and mass spectrometry while in present study used RT-PCR because it is more sensitive and provide quantitative measurement of RNA level, Annexin A2 is present in the cytoplasm and on cell surfaces this gene can bind membranes to other membranes for reason called bridge (Annexin A2) [23],during inflammation ANNEXIN A2 modulates the nuclear factor kappa-B (NF-κB) and cell apoptosis signaling pathways and have role in guide



the chemotaxis of inflammatory cells toward inflammation sites Which contributes to increased inflammation [24] inflammatory cell secretion in oviduct It has a negative effect on oviduct environment and effect on fertilization process ,Annexin A2 role chemotaxis attract of pathogen to site infection facilitate their invasion and proliferation , the female genital contain bacteria also in oviduct [25,26,27] , role in proliferation of this bacteria and increase infection ,inflammation reduce oocyte quality and causes damage in zona pellucida [28,29] impair oocyte wall and chemotaxis effect of ANNEXIN A2 can contribute attract of sperm toward oocyte lead to polyspermy and failure of fertilization , [30]and [31] found that ANNEXIN A2 in human and mice can contribute causes fibrosis in lung and liver according to that it may be have the same role in the oviduct fibrosis formation lead to obstruction and impair oocyte and sperm movement through oviduct and possible permanent infertility when Annexin a2 increase, the increase gene expression of ANNEXIN A2 in the oviduct is affected by endometritis and that inflammation has extended to oviduct ampulla, the expression of ANNEXIN A2 acts as a double edged sword it crucial role for sperm-oocyte adhesion and enhances fertilization rate , and the presence of inflammation it turns to negative effect by attracting inflammatory cell to site of infection and lead to chronic inflammation.

Conclusion

The presence of macroscopic lesion in the uterus does not necessarily as indicator for microscopic pathological change. Increased expression of the Annexin A2 in oviduct ampulla in cows with endometritis. Endometritis extends its effects on the oviduct by it is effect on the oviduct genes.

Acknowledgment

The authors would like to acknowledge the College of Veterinary Medicine, University of Al-Qadisiyah, Al-Diwanyah, Iraq, for the support.

Conflict of interest

No conflict of interest is found for the present study.

Funding source

This research had no specific fund; however, it was self-funded by the authors.

References

- 1.Tucho TT, Ahmed WM. Economic and reproductive impacts of retained placenta in dairy cows. *Journal of Reproduction and Infertility.* 2017;8(1):18-27.
- 2.Azawi OI, Omran SN, Hadad JJ. A study of endometritis causing repeat breeding of cycling Iraqi

buffalo cows. *Reproduction in Domestic Animals.* 2008 Dec;43(6):735-43.

- 3.Pascottini OB, LeBlanc SJ, Gnemi G, Leroy JL, Opsomer G. Genesis of clinical and subclinical endometritis in dairy cows. *Reproduction.* 2023 Aug 1;166(2):R15-24.

- 4.Bastos NM, Ferst JG, Goulart RS, Coelho da Silveira J. The role of the oviduct and extracellular vesicles during early embryo development in bovine. *Animal Reproduction.* 2022 Apr 20;19(1):e20220015.

- 5.Avilés M, Coy P, Rizos D. The oviduct A key organ for the success of early reproductive events. *Departamento de Reproducción Animal.* 2015.

- 6.Miller DJ. Regulation of sperm function by oviduct fluid and the epithelium: insight into the role of glycans. *Reproduction in domestic animals.* 2015 Jul;50:31-9.

- 7.VR A, KM L. structural defects of the oviduct causing infertility in crossbred dairy cows. *Journal of Indian Veterinary Association.* 2022 Apr 1;20(1).

- 8.Teijeiro JM, Roldán ML, Marini PE. Annexin A2 and S100A10 in the mammalian oviduct. *Cell and Tissue Research.* 2016 Feb;363:567-77.

- 9.Almiñana C, Corbin E, Tsikis G, Alcántara-Neto AS, Labas V, Reynaud K, Galio L, Uzbekov R, Garanina AS, Druart X, Mermillod P. Oviduct extracellular vesicles protein content and their role during oviduct–embryo cross-talk. *Reproduction.* 2017 Sep 1;154(3):253-68.

- 10.Dallacasagrande V, Hajjar KA. Annexin A2 in inflammation and host defense. *Cells.* 2020 Jun 19;9(6):1499.

- 11.Lim HI, Hajjar KA. Annexin A2 in fibrinolysis, inflammation and fibrosis. *International Journal of Molecular Sciences.* 2021 Jun 25;22(13):6836.

- 12.He X, Zhang W, Chang Q, Su Z, Gong D, Zhou Y, Xiao J, Drelich A, Liu Y, Popov V, Zhao X. A new role for host annexin A2 in establishing bacterial adhesion to vascular endothelial cells: lines of evidence from atomic force microscopy and an in vivo study. *Laboratory Investigation.* 2019 Nov 1;99(11):1650-60.

- 13.Kuehnl A, Musiol A, Raabe CA, Rescher U. Emerging functions as host cell factors—an encyclopedia of annexin-pathogen interactions. *Biological Chemistry.* 2016 Oct 1;397(10):949-59.

- 14.Lopera-Vásquez R, Uribe-García F, Rondón-Barragán I. Effect of estrous cycle phases on gene expression in bovine oviduct epithelial cells. *Veterinary World.* 2022 Jul 14;15(7):1665.

- 15.Rhyaf AG. Histopathological study of endometritis of the cows. *Al-Qadisiya J Vet Med Sci.* 2010;9(1):1-6.



16. Al-Attar A, Al-Allaf SM, Al-Mokhtar KA. Microscopic preparations.. National Library in Baghdad. Iraq. Bernard. 1982.
17. Noory MA. Science Technology microscopic slides. Mosul University.
18. Ghanem N, Hölker M, Rings F, Jennen D, Tholen E, Sirard MA, Torner H, Kanitz W, Schellander K, Tesfaye D. Alterations in transcript abundance of bovine oocytes recovered at growth and dominance phases of the first follicular wave. *BMC Developmental Biology*. 2007 Dec;7:1-9.
19. Schmittgen TD, Livak KJ. Analyzing real-time PCR data by the comparative CT method. *Nature protocols*. 2008 Jun;3(6):1101-8.
20. Suleymanov SM, Usha BV, Vatnikov YA, Sotnikova ED, Kulikov EV, Parshina VI, Bolshakova MV, Lyshko MU, Romanova EV. Structural uterine changes in postpartum endometritis in cows. *Veterinary World*. 2018 Oct 22;11(10):1473.
21. Al-delemi. D.H.J. Anatomical, Physiological, Bacteriological & Pathological Study of The Reproductive System of Iraqi She-Camels (Camelus dromedaries) .Ph.D. Thesis , College of Veterinary Medicine, University of Baghdad (2007).
22. Ledgard AM, Smolenski GA, Henderson H, Lee RF. Influence of pathogenic bacteria species present in the postpartum bovine uterus on proteome profiles. *Reproduction, Fertility and development*. 2015 Feb 13;27(2):395-406.
23. Gerke V, Creutz CE, Moss SE. Annexins: linking Ca²⁺ signalling to membrane dynamics. *Nature reviews Molecular cell biology*. 2005 Jun 1;6(6):449-61.
24. Wang T, Zhao D, Zhang Y, Yu D, Liu G, Zhang K. Annexin A2: A Double-Edged Sword in Pathogen Infection. *Pathogens*. 2024 Jul 4;13(7):564.
25. Al-delemi, D.H. & Al-hilali, H.A. The Uterine Bacterial Flora of Normal Reproductive Tract ,Non – Pregnant Iraqi Cows. *The Veterinarian, Iraqi Veterinary Medicine Syndicate*. (2001). Vol.11, No.1.
26. Al-delemi, D.H., Abo-Dekhen, S.K. and Al-hilali, H.A. The normal bacterial flora in the uteri of Iraqi pregnant cows *AL-Qadisiya Journal of Veterinary Medicine Science*. (2003). .Vol.2 . No.1 .
27. Garcia-Garcia RM, Arias-Álvarez M, Jordán-Rodríguez D, Rebollar PG, Lorenzo PL, Herranz C, Rodríguez JM. Female reproduction and the microbiota in mammals: Where are we?. *Theriogenology*. 2022 Dec 1;194:144-53.
28. Calongos G, Hasegawa A, Komori S, Koyama K. Harmful effects of anti-zona pellucida antibodies in folliculogenesis, oogenesis, and fertilization. *Journal of Reproductive Immunology*. 2009 Jan 1;79(2):148-55.
29. Snider AP, Wood JR. Obesity induces ovarian inflammation and reduces oocyte quality. *Reproduction*. 2019 Sep 1;158(3):R79-90.
30. Lei Y, Wang K, Li X, Li Y, Feng X, Zhou J, Zhang Z, Huang C, Zhang T. Cell-surface translocation of annexin A2 contributes to bleomycin-induced pulmonary fibrosis by mediating inflammatory response in mice. *Clinical Science*. 2019 Apr;133(7):789-804.
31. Zhang L, Peng X, Zhang Z, Feng Y, Jia X, Shi Y, Yang H, Zhang Z, Zhang X, Liu L, Yin L. Subcellular proteome analysis unraveled annexin A2 related to immune liver fibrosis. *Journal of cellular biochemistry*. 2010 May 1;110(1):219-28.