

The comparative Effects of Nano and Conventional Spirulina Extracts on Oxidative Stress and Hepatic Biomarkers in Hyperlipidemic male Rats

Fikra muhammed mansi* Hussein Abbas salman

Department of Biology, College of Education, University of Al-Qadisiyah, Iraq

*Corresponding Author email: edu.bio.postal16@qu.edu.iq

Submitted: May 04, 2025

Revised: May 26, 2025

Accepted: June 01, 2025

Correspondence

Fikra muhammed mansi
edu.bio.postal16@qu.edu.iq

Abstract Obesity is a complicated metabolic disorder characterized by abnormal or excessive fat accumulation in the body that may pose a higher risk for chronic diseases. The present study investigates the protective effects of *Spirulina platensis* extracts (SPEs) from both conventional and nano-formulated extraction against oxidative stress and liver function in rats with hyperlipidemia. Forty male albino rats were divided into five groups. The C group received normal saline only during the observation period. Hyperlipidemic group (T1) was treated with high fat diet (HFD) for 90 days without any medication. Group T2 was treated with the conventional ethanolic *Spirulina* extract (100 mg/kg/day) during the HFD. Group T3 was also administered *Spirulina* extract in a nano-form and in the same dose. Group T4 treated with ZnO-NPs (10 mg/kg/day) with HFD. Animals were treated for 30 days after hyperlipidemia induction. The body weight, and serum MDA, liver enzymes (ALT and AST) were significantly increased while antioxidant enzymes (GSH, CAT and SOD) were prominently reduced in T1 group, indicating oxidative damage. *Spirulina* treatment, particularly in nano-form (T3), resulted in remarkable enhancement in all previously evaluated parameters. The T3 treatment had the highest increment effect followed by the T2 and T4. This study concluded that nano-formulated *Spirulina* (T3) showed higher protection level compared to conventional form (T2) and ZnO-NPs (T4) implying that nanotechnology increases biological activity of natural antioxidants, especially under conditions associated with lipid accumulation and development of oxidative stress.

Keywords: *Spirulina platensis*; oxidative stress; liver enzymes; hyperlipidemia; antioxidant biomarkers.

©Authors, 2025, College of Veterinary Medicine, University of Al-Qadisiyah. This is an open access article under the CC BY 4.0 license (<http://creativecommons.org/licenses/by/4.0/>).

Introduction Hyperlipidemia is defined as an abnormally high concentration of lipids and lipoproteins in the blood. It is a group of heterogeneous disorders that result in the formation of a group of excess lipids in the bloodstream due to an increase in one of the types of lipids, either hypercholesterolemia, hypertriglyceridemia, or both (mixed hyperlipidemia). This increase is associated with the development of atherosclerosis, which is the main cause of coronary heart disease, ischemic heart disease, stroke, and kidney failure, which have witnessed a significant increase in our time (1).

Chronic administration of a high-fat diet (HFD) is also one of the most prevalent triggers of lipid imbalance and oxidative stress in mammals, which can lead to lipid peroxidation, systemic inflammation, and damage to multiple organs (2, 3). Recently, attention has been directed towards natural bioactive compounds that can counter lipid abnormalities. Among them, *Spirulina platensis*, a blue-green alga

(Cyanobacteria), has attracted special attention due to its very promising nutritional and medicinal potential. It contains several antioxidants, vitamins (B-complex, C, E), minerals (iron, magnesium, selenium), essential amino acids, and bioactive pigments like phycocyanin and β -carotene (4). Reports indicate *Spirulina* has antioxidant, anti-inflammatory, hypolipidemic, immunostimulatory, and hepatoprotective properties (5, 6). Inclusion of *Spirulina* in nano-formulations to enhance its bioactivity has also been suggested. With large surface area and bioavailability, nanoparticles encourage efficient cellular absorption and targeted delivery. Specialized nanomaterials such as Zinc Oxide Nanoparticles (ZnO-NPs) have shown potential therapeutic effects due to the antioxidative impacts *Spirulina* remain scarce (7). The objective of this study was to investigate the comparative physiological and biochemical impacts of Ethanolic extract of *Spirulina platensis*, its nanoform and Nano-

ZnO on hyperlipidemia induced by high fat diet in male rats. The evaluated parameters were as follows: body weight gain, oxidative stress indicators (MDA, GSH, CAT, SOD) and liver functions enzymes (ALT, AST).

Material and Methods

Ethical Approval Statement

This work was accepted by the Medical Ethics Committee, College of Education, Al-Qadisiyah University. All animal studies were performed in compliance with the institutional guidelines and ethical standards. Written and oral informed consent were sought from the responsible academic quarters before the outset of the study.

Experimental Design

The rats were randomly divided, the first group (G1, control group) consisted of eight rats administered normal saline (0.9% NaCl) throughout the duration of the experiment. The second group (G2) included thirty-two rats subjected to a high-fat diet (HFD) supplemented with 30% animal fat for 60 days to induce hyperlipidemia, as described by (8). The animals in G2 were further subdivided into four treatment groups (n=8 per group) as follows:

- T1 (Positive Control Group): Received only the high-fat diet (30% animal fat) for an additional 30 days without any treatment.
- T2: Administered ethanolic extract of *Spirulina platensis* at a dose of 100 mg/kg body weight via oral gavage daily for 30 days, while continuing on the high-fat diet.
- T3: Received nano-formulated ethanolic extract of *Spirulina platensis* at the same dose (100 mg/kg) under identical dietary conditions.
- T4: Treated with zinc oxide nanoparticles (ZnO NPs) at a dose of 10 mg/kg body weight via oral gavage daily while maintained on the high-fat diet.

Chemicals and Dose Justification

The *Spirulina* powder was obtained from a certified local supplier and taxonomically authenticated by specialists in the Department of Biology, University of Al-Qadisiyah. Zinc oxide nanoparticles were acquired and validated using Fourier-transform infrared spectroscopy (FTIR) in the Chemistry Department of the same university.

The dose for the ethanolic extract of *Spirulina platensis* was selected based on prior studies that demonstrated its physiological efficacy at 100 mg/kg (9, 10). The same dose was applied to the nano-formulated extract. The ZnO NPs dose (10 mg/kg) was determined according to (11). All treatments were administered once daily via oral gavage for a period of 30 days.

Preparation of Conventional and Nano-formulated Ethanolic Extracts

The *Spirulina* powder was finely ground using an electric mill. The ethanolic extract was prepared following the method outlined by (12), while the nano-formulated extract was synthesized by loading the ethanolic extract onto nanoparticle carriers based on the method reported by (13). The prepared formulations were stored in sterile, sealed plastic containers and used immediately for dosing.

Assessment of Oxidative Stress Markers

Blood samples were collected at the end of the experiment via cardiac puncture under ketamine/xylazine anesthesia. Serum was isolated and used to evaluate oxidative stress biomarkers. Malondialdehyde (MDA) levels were estimated using a modified method described by Guidet and Shah (14). Levels of reduced glutathione (GSH), catalase (CAT), and superoxide dismutase (SOD) were measured using protocols by (15), (16), and (17), respectively.

Liver Enzyme Activity

Serum levels of alanine aminotransferase (ALT) and aspartate aminotransferase (AST) were determined using the colorimetric method of Reitman and Frankel (18) to assess hepatic function.

Body Weight Gain Calculation

Body weight gain was assessed by calculating the difference between the final and initial body weights of the animals at the end of the experimental period using the formula:

$$\text{Weight Gain (g)} = \text{Final Body Weight} - \text{Initial Body Weight}$$

Statistical Analysis

All data were analyzed using SPSS version 27. One-way analysis of variance (ANOVA) was performed to evaluate differences among groups, followed by the Least Significant Difference (LSD) test to identify statistically significant pairwise comparisons. Results were considered statistically significant at $p \leq 0.05$ (19).

Results and discussion

Body Weight Gain

The results shown in Table (1) reflect the impact of different treatments on the body weight gain of rats exposed to a high-fat diet. The T1 group, which continued to receive the fat-enriched feed without any therapeutic intervention, recorded a significantly high weight gain (208.5 ± 14.05 g), confirming the successful induction of hyperlipidemia. This value was substantially higher than that of the negative control group (C), which recorded a normal physiological weight gain of 97.25 ± 7.55 g. On the other hand, treatment with either the conventional ethanolic extract (T2) or the nano-

formulated extract (T3) of *Spirulina platensis* led to a significant reduction in body weight gain, registering 52.5 ± 15.63 g and 30.25 ± 3.01 g respectively. These findings suggest that *Spirulina* in both forms has an anti-obesogenic effect, with the nano-form being more effective. Additionally, the group treated with zinc oxide nanoparticles (T4) also showed a significant decline in weight gain (79 ± 22.64 g), although this reduction was less pronounced than that observed in the *Spirulina*-treated groups.

Table 1. Effect of Different Treatments on Body Weight Gain in Experimental Rat Groups (Mean \pm SD)

Group	Initial Weight (g)	Final Weight (g)	Weight Gain (g)
C	259 \pm 3.18 (a)	356.25 \pm 4.71 (a)	97.25 \pm 7.55 (b)
T1	240 \pm 17.19 (a)	448.5 \pm 3.86 (a)	208.5 \pm 14.05 (a)
T2	250.25 \pm 10.11 (a)	30.75 \pm 20.29 (d)	52.5 \pm 15.6 3(d)
T3	264 \pm 6.79 (a)	294.25 \pm 6.10 (e)	30.25 \pm 3.01 (e)
T4	249.25 \pm 5.39 (a)	328.25 \pm 19.01 (c)	79 \pm 22.64 (c)
LSD	15.214	16.254	11.241

Note: Small Different letters indicate statistically significant differences between treatments $p \leq 0.05$

The numbers indicate the mean \pm standard error

C: The control group was dosed with physiological saline for the duration of the experiment .

T1: The first treatment represents the group of rats in which Hyperlipidemia induced by feeding on high fat diet.

T2: The second treatment represents the group of rats that were dosed with alcoholic extract of *S. Platensis* for the duration of the experiment(100ml/kg).

T3: The third treatment represents the group of rats that were dosed with the Nanoalcoholic extract of *S. Platensis* for the length of the experiment(100ml/kg).

T4: The third treatment represents the group of rats that were dosed with nano-zinc oxide for the duration of the experiment(10 ml/kg).

The statistically significant difference ($p \leq 0.05$) among the groups illustrates that dietary intervention with *Spirulina platensis*, especially in nano-form, effectively suppresses body weight gain in hyperlipidemic rats. The high-fat diet in T1 likely enhanced bile secretion, lipid absorption, and triglyceride deposition in tissues, aligning with prior studies that demonstrated the obesogenic impact of cholesterol- and cholic-acid-enriched diets (20–23).

The anti-obesity effects observed in the T2 and T3 groups can be attributed to *Spirulina*'s rich nutritional profile, including amino acids, vitamins, and high-quality proteins superior even to soybean (24, 25).

One suggested mechanism involves L-phenylalanine-induced stimulation of cholecystokinin, which suppresses appetite (26). Clinical findings also confirm *Spirulina*'s weight-reducing and metabolic benefits in humans (27, 28).The more pronounced effect in the T3 group indicates that nano-formulated *Spirulina* enhances bioavailability and interaction with cellular receptors due to increased surface area, consistent with findings from (29, 30).T4's moderate weight suppression effect may relate to zinc's role in insulin sensitivity and metabolic regulation, although it was less effective than *Spirulina* (31).

Oxidative Stress Biomarkers

Table 2 displays the serum levels of oxidative stress markers. The T1 group exhibited significantly elevated levels of malondialdehyde (MDA: 4.86 ± 0.36 mmol/L), a marker of lipid peroxidation, indicating oxidative damage due to the high-fat diet. Conversely, antioxidant enzyme levels—glutathione (GSH), catalase (CAT), and superoxide dismutase (SOD)—were significantly lower in T1 compared to the control group.

Treatment with *Spirulina* (T2 and T3) and ZnO NPs (T4) reversed these alterations. The nano-form (T3) showed the highest improvements in antioxidant parameters: GSH (16.26 ± 1.93), CAT (0.500 ± 0.01), and SOD (2.02 ± 0.05), while reducing MDA to 3.16 ± 0.09 .

Table 2. Oxidative Stress Related Biomarkers (GSH, CAT, SOD, and MDA) in Serum Levels

Group	GSH (mmol/L)	CAT (mmol/L)	SOD (mmol/L)	MDA (mmol/L)
C	22.78 \pm 1.2 (3 a)	0.730 \pm 0.0 3 (a)	3.16 \pm 0.11 (a)	2.35 \pm 0.0 5 (e)
T1	9.15 \pm 0.15 (d)	0.207 \pm 0.0 1 (e)	1.17 \pm 0.06 (e)	4.86 \pm 0.3 6 (a)
T2	13.59 \pm 2.0 7 (c)	0.332 \pm 0.0 3 (c)	1.66 \pm 0.07 (c)	3.61 \pm 0.2 8 (e)
T3	16.26 \pm 1.9 3 (b)	0.500 \pm 0.0 1 (b)	2.02 \pm 0.05 (b)	3.16 \pm 0.0 (d) 9
T4	15.03 \pm 0.9 4 (bc)	0.290 \pm 0.0 1 (d)	1.40 \pm 0.03 (d)	3.99 \pm 0.0 7 (b)
LSD	2.18	0.038	0.116	0.327

Note: Small Different letters indicate statistically significant differences between treatments $p \leq 0.05$

The numbers indicate the mean \pm standard error

C: The control group was dosed with physiological saline for the duration of the experiment.

T1: The first treatment represents the group of rats in which Hyperlipidemia induced by feeding on high fat diet.

T2: The second treatment represents the group of rats that were dosed with alcoholic extract of *S. Platensis* for the duration of the experiment(100ml/kg).

T3: The third treatment represents the group of rats that were dosed with the Nanoalcoholic extract of *S. Platensis* for the length of the experiment(100ml/kg).

T4: The third treatment represents the group of rats that were dosed with nano-zinc oxide for the duration of the experiment (10 ml/kg).

The (T1) group's oxidative imbalance may stem from cholesterol-induced ROS generation and subsequent cytokine-mediated neutrophil activation (34). Lipid peroxidation impairs cell membranes, depletes GSH, and reduces enzymatic antioxidant defenses (35). This is consistent with reports from (34, 35), who observed MDA elevation and SOD/CAT suppression in hyperlipidemic rats. The restoration of antioxidant levels in *Spirulina*-treated groups likely results from phycocyanin and carotenoids, which act as potent free radical scavengers and anti-inflammatory agents (36). T3's superior effect reaffirms the therapeutic promise of nano-delivery systems for enhancing cellular uptake (37).

Table (3) illustrates serum transaminase levels. The T1 group showed significantly elevated ALT (91.07 ± 7.78 U/L) and AST (105.23 ± 6.73 U/L), reflecting hepatic injury due to prolonged lipid overload. The control group recorded physiological levels (ALT: 33.41 ± 6.07 ; AST: 43.07 ± 4.77).

In contrast, T2, T3, and T4 groups demonstrated significant reductions, with the T3 group exhibiting the most pronounced improvement: ALT (53.82 ± 5.89), AST (58.57 ± 8.13). T2 and T4 followed in efficacy.

Table (3). Effect of Different Treatments on Serum AST and ALT Levels in Experimental Rat Groups (Mean \pm SD)

Group	ALT (U/L)	AST (U/L)
C	(33.41 ± 6.07) e	(43.07 ± 4.77) d
T1	(91.07 ± 7.78) a	(105.23 ± 6.73) a
T2	(60.76 ± 0.70) c	(77.94 ± 0.7) b
T3	(53.82 ± 5.89) d	(58.57 ± 8.13) c
T4	(60.76 ± 0.70) b	(77.94 ± 0.7) b
LSD	8.23	10.16

Note: Small Different letters indicate statistically significant differences between treatments $p \leq 0.05$. The numbers indicate the mean \pm standard error

C: The control group was dosed with physiological saline for the duration of the experiment.

T1: The first treatment represents the group of rats in which Hyperlipidemia induced by feeding on high fat diet.

T2: The second treatment represents the group of rats that were dosed with alcoholic extract of *S. Platensis* for the duration of the experiment(100ml/kg).

T3: The third treatment represents the group of rats that were dosed with the Nanoalcoholic extract of *S. Platensis* for the length of the experiment(100ml/kg).

T4: The third treatment represents the group of rats that were dosed with nano-zinc oxide for the duration of the experiment (10 ml/kg).

ALT and AST elevation in T1 reflects hepatocellular membrane damage, enzyme leakage, and hepatic oxidative burden (42–44). Improvement in the *Spirulina*-treated groups likely stem from the stabilization of hepatocyte membranes and reduction of inflammatory mediators, as documented in prior studies (38).

Mazokopakis et al. demonstrated that oral *Spirulina* intake lowered liver enzymes in NAFLD patients (39), and similar effects were observed in animal studies (40, 41). The use of nano-formulation in T3 enhances *Spirulina*'s hepatic delivery and therapeutic response, consistent with the benefits of nanoparticle-mediated targeting (42).

Conclusion

The present study proved that the normal and nano-ethanolic extracts of *Spirulina platensis* ameliorate the body weight, antioxidant capacity and liver functions in hyperlipidemic rats. Regarding both forms, the nano-form exhibited better protection against weight gain, induction of antioxidant enzymes, and regulation of liver markers. These findings highlight the higher efficacy of nanotechnology in enhancing the therapeutic effectiveness of *Spirulina* in metabolic diseases, such as obesity and hepatic diseases.

Acknowledgement

The Authors are highly thankful to the Department of Biology- College of Pedagogy -University of AL-Qadisiyah for the provision of lab facilities and technical assistance throughout the present study. We would like to thank all of the staff at the Animal House for their help with restraining and husbandry of the laboratory animals.

Funding

The study was self-funded by the authors.

Conflict of interest

No conflict of interest is disclosed by the authors.

References

- Huang CJ, McAllister MJ, Slusher AL, Webb HE, Mock JT, Acevedo EO. Obesity-related oxidative stress: the impact of physical activity and diet manipulation. *Sports medicine-open*. 2015 Dec;1:1-2.
- Mukhopadhyay SK. Study of lipid profile in obese individuals and the effect of cholesterol lowering agents on them. *A meen J Med Sci*. 2012 Jan 1;5(2):147-51.

- 3-Noria SF, Grantcharov T. Biological effects of bariatric surgery on obesity-related comorbidities. *Canadian Journal of Surgery*. 2013 Feb;56(1):47.
- 4-Priyadarshani I, Rath B. Commercial and industrial applications of micro algae—A review. *Journal of Algal Biomass Utilization*. 2012 Oct;3(4):89-100.
- 5-Koru E. Earth food Spirulina (Arthrospira): production and quality standards. *Food additive*. 2012 Feb 22;10:31848.
- 6-Verma AK, Dewangan K, Daunday L, Naurange K, Verma K, Bhiaram M. Spirulina as functional food: insights into cultivation, production, and health benefits. *Journal of Applied Pharmaceutical Research*. 2024 Oct 31;12(5):28-50.
- 7-Karkos PD, Leong SC, Karkos CD, Sivaji N, Assimakopoulos DA. Spirulina in clinical practice: evidence-based human applications. *Evidence Based Complementary and Alternative Medicine*. 2008;2011:27.
- 8-Altunkaynak Z. Effects of high fat diet induced obesity on female rat livers (a histochemical study). *European Journal of General Medicine*. 2005 Jul 15;2(3):100-9.
- 9-Al-Gabri DT, Al-Naely AJ, Alghanmi HA. Using of nanocomposite loading *klisinema persicum* for reducing the damage of the liver and kidneys in female rats caused by taxol (paclitaxel). *Turkish Journal of Physiotherapy and Rehabilitation*. 2021;32(3).
- 10-Mawed SA, Centoducati G, Farag MR, Alagawany M, Abou-Zeid SM, Elhady WM, El-Saadony MT, Di Cerbo A, Al-Zahaby SA. *Dunaliella salina* microalga restores the metabolic equilibrium and ameliorates the hepatic inflammatory response induced by zinc oxide nanoparticles (ZnO-NPs) in male zebrafish. *Biology*. 2022 Oct 1;11(10):1447.
- 11-Djearmane S, Lim YM, Wong LS, Lee PF. Cytotoxic effects of zinc oxide nanoparticles on cyanobacterium *Spirulina (Arthrospira) platensis*. *PeerJ*. 2018 Jun 1;6:e4682.
- 12-Gebrehiwot S, Giday M, Erko B, Mekonnen Y. In vivo antimalarial activity of *Capparis tomentosa* Lam. in mice infected with *Plasmodium berghei*. *Journal of Ayurvedic and Herbal Medicine*. 2019;5(2):44-8.
- 13-Karkos PD, Leong SC, Karkos CD, Sivaji N, Assimakopoulos DA. Spirulina in clinical practice: evidence-based human applications. *Evidence-based complementary and alternative medicine*. 2011;2011(1):531053.
- 14-Ibrahim DA, Zbbar SA, Salih MA. Evaluation the Role of Malondialdehyde in Myocardial Infarction Patients. *Indian Journal of Forensic Medicine & Toxicology*. 2021 Oct 1;15(4).
- 15-Sedlak J, Lindsay RH. Estimation of total, protein-bound, and nonprotein sulfhydryl groups in tissue with Ellman's reagent. *Analytical biochemistry*. 1968 Jan 1;25:192-205.
- 16-Mueller S, Riedel HD, Stremmel W. Direct evidence for catalase as the predominant H₂O₂-removing enzyme in human erythrocytes. *Blood, The Journal of the American Society of Hematology*. 1997 Dec 15;90(12):4973-8.
- 17-Durak I, Canbolat O, Kavutçu M, Öztürk HS, Yurtarslani Z. Activities of total, cytoplasmic, and mitochondrial superoxide dismutase enzymes in sera and pleural fluids from patients with lung cancer. *Journal of clinical laboratory analysis*. 1996;10(1):17-20.
- 18-Clermont RJ, Chalmers TC. The transaminase tests in liver disease. *Medicine*. 1967 Mar 1;46(2):197-207.
- 19-Ab Rahman J. Brief guidelines for methods and statistics in medical research. Springer Singapore; 2015 Oct 14.
- 20-El-Sheekh MM, Hamad SM, Goma M. Protective effects of Spirulina on the liver function and hyperlipidemia of rats and human. *Brazilian archives of Biology and Technology*. 2014;57:77-86.
- 21-Sarumathi A, Sethupathy S, Saravanan N. The protective efficacy of spirulina against bacterial endotoxin potentiated alcoholic liver disease. *Journal of Functional Foods*. 2014 Jul 1;9:254-63.
- 22-Khafaga AF, El-Sayed YS. Spirulina ameliorates methotrexate hepatotoxicity via antioxidant, immune stimulation, and proinflammatory cytokines and apoptotic proteins modulation. *Life sciences*. 2018 Mar 1;196:9-17.
- 23-Bandarrigi M, Shakerian S, Ranjbar R, Abdollahi S. The Effect of Eight Weeks of Yoga Practice with Weight along with Spirulina Supplement on Some Indicators of Metabolic Syndrome in Obese and Overweight Older Women. *Journal of Nutrition, Fasting & Health*. 2023 Jul 1;11(3).
- 24-Li X, Liang S, Tan CH, Cao S, Xu X, Er Saw P, Tao W. Nanocarriers in the enhancement of therapeutic efficacy of natural drugs. *BIO Integration*. 2021 Jul 1;2(2):40.
- 25-Bitam A, Aissaoui O. Spirulina platensis, oxidative stress, and diabetes. In *Diabetes 2020* Jan 1 (pp. 325-331). Academic Press.

- 26-Ross AC, Harrison EH. Vitamin A: nutritional aspects of retinoids and carotenoids. *Handbook of vitamins*. 2007 Jun 8;4:1-39.
- 27-Kaps L, Limeres MJ, Schneider P, Svensson M, Zeyn Y, Fraude S, Cacicedo ML, Galle PR, Gehring S, Bros M. Liver cell type-specific targeting by nanoformulations for therapeutic applications. *International Journal of Molecular Sciences*. 2023 Jul 24;24(14):11869.
- 28-Ipar VS, Dsouza A, Devarajan PV. Enhancing curcumin oral bioavailability through nanoformulations. *European journal of drug metabolism and Pharmacokinetics*. 2019 Aug 1;44:459-80.
- 29-Iravani S, Varma RS. Advanced drug delivery micro-and nanosystems for cardiovascular diseases. *Molecules*. 2022 Sep 9;27(18):5843.
- 30-Arrari F, Jabri MA, Ayari A, Dakhli N, Ben Fayala C, Boubaker S, Sebai H. Chromatographic analyses of spirulina (*arthrospira platensis*) and mechanism of its protective effects against experimental obesity and hepatic steatosis in rats. *Medicina*. 2023 Oct 13;59(10):1823.
- 31-Manzoni AG, Passos DF, Leitemperger JW, Storck TR, Doleski PH, Jantsch MH, Loro VL, Leal DB. Hyperlipidemia-induced lipotoxicity and immune activation in rats are prevented by curcumin and rutin. *International immunopharmacology*. 2020 Apr 1;81:106217.
- 32-Abela GS, Vedre A, Janoudi A, Huang R, Durga S, Tamhane U. Effect of statins on cholesterol crystallization and atherosclerotic plaque stabilization. *The American journal of cardiology*. 2011 Jun 15;107(12):1710-7.
- 33-Shantha NC, Napolitano GE. Gas chromatography of fatty acids. *Journal of Chromatography A*. 1992 Oct 30;624(1-2):37-51.
- 34-Lopez CA, Beavers WN, Weiss A, Knippel RJ, Zackular JP, Chazin W, Skaar EP. The immune protein calprotectin impacts *Clostridioides difficile* metabolism through zinc limitation. *MBio*. 2019 Dec 24;10(6):10-128.
- 35-Wu Q, Liu L, Miron A, Klímová B, Wan D, Kuča K. The antioxidant, immunomodulatory, and anti-inflammatory activities of Spirulina: an overview. *Archives of toxicology*. 2016 Aug;90:1817-40.
- 36-Diraman H, Koru E, Dibeklioglu H. Fatty acid profile of *Spirulina platensis* used as a food supplement. *Israeli Journal of Aquaculture-Bamidgeh*. 2009 Jan 1;61.
- 37-El-Demerdash, F. M., et al. (2005). Onion and garlic as hypoglycemic agents. *Food and Chemical Toxicology*, 43(1), 57–63.
- 38-Faraji S, Daneghian S, Alizadeh M. Effects of chicory (*Cichorium intybus* L.) on nonalcoholic fatty liver disease. *Traditional Medicine Research*. 2020 Aug 31;5(6):476-86.
- 39-El-Boghdady NA, Kamel MA, El-Shamy RM. Omeprazole and spirulina platensis ameliorate steatohepatitis in experimental nonalcoholic fatty liver disease. *Metabolic Syndrome and Related Disorders*. 2020 Nov 1;18(9):426-34.
- 40-Shi M, Chen S, Feng Y, Wang S, Xia Y, He J. Marine natural products as an important source of bioactive substances for non-alcoholic fatty liver disease management. *Frontiers in Marine Science*. 2025 Jan 28;11:1523246.
- 41-Mansour SA, Mossa AT. Lipid peroxidation and oxidative stress in rat erythrocytes induced by chlorpyrifos and the protective effect of zinc. *Pesticide biochemistry and physiology*. 2009 Jan 1;93(1):34-9.
- 42-Prasad AS. Discovery of human zinc deficiency: 50 years later. *Journal of Trace Elements in Medicine and Biology*. 2012 Jun 1;26(2-3):66-9.