



The Sheep Pox Virus's Antiapoptotic Gene Sequence Analysis as a Host Immunity Evading Gene

Rasha Hussain Mohammed Al-Saadi[✉]

¹Department of Internal and Preventive Medicine, College of Veterinary Medicine, University of Al-Qadisiyah, Iraq

<https://orcid.org/0000-0003-2751-862X>[✉]

Submitted: July 25, 2025

Revised: August 01, 2025

Accepted: August 09, 2025

Correspondence

Rasha Hussain

vet.post22.21@qu.edu.iq

Abstract Sheep pox is a highly contagious infectious disease that primarily affects small ruminants such as sheep. The causative agent is a member of the Poxviridae family, and causes severe skin rashes, fever, diarrhea, deaths in young litters and abortions in pregnant ewes. In recent years, many single and multiple cases of SPV have been reported with failure of control and treatment strategies. This indicates that the SPV virus may undergo genetic changes that reduce host immunity, so it had to be studied to diagnose the gene encoded by the virus and identify its sequences, such as the apoptosis gene. One hundred and twenty-five scab samples were collected from suspected in the Diwaniyah Province throughout September 2023 and January 2024, the samples were subjected to a PCR examination to identify the presence of the anti-apoptosis gene. Result revealed that the expected amplicons size 515 bp of SPV was detected in forty samples (32%) of skin lesions. Number of base substitutions per site between sequences was shown. The identity score of apoptosis related gene of ten positive local isolates was 100% with Abu Gharib_Iraq vaccines Registered breeds in global sequences. Phylogenetic tree analysis based on the partial apoptosis related gene showed that all samples tested were closely related in sequence alignment with NCBI-BLAST capripoxviruses: sheeppox virus envelope protein apoptosis related gene. This study provided sequence information of anti-apoptotic gene for several SPV isolates, which positively affects the epidemiological study of Capripoxvirus.

Keywords: apoptosis related genes, Capripoxvirus, PCR, SPV.

©Authors, 2025, College of Veterinary Medicine, University of Al-Qadisiyah. This is an open access article under the CC BY 4.0 license (<http://creativecommons.org/licenses/by/4.0/>).

Introduction Sheeppox virus (SPV) is an international viral disease that primarily affects sheep and goats, causing skin lesions and fever (1,2,3). Causing large losses in countries with extensive small ruminant breeding programs (4). This disease imposes significant economic costs and risks on the country, neighbors and trading partners, Trade bans can be mentioned due to the presence of these major cross-border diseases (5,6). The virus responsible for sheep pox and goat pox is Capri poxvirus, a large double-stranded DNA virus with dimensions of 170-260 nm by 300-450 nm from the poxviridae family (7,8). This enormous genome encodes every gene required for their specific intracellular replication (9). Clinical indicators can include fever, nodules and papules, internal sores in the lungs, respiratory, and gastrointestinal mucosa, and cause the animals' mortality (10,11,12). Other symptoms include decreased milk yield and weight gain, higher rates of miscarriage, greater susceptibility to pneumonia, and high mortality (13,14). The geographical distribution of sheep pox has been rather steady. Sheep pox and

goat pox are widespread in many countries, including Iraq, Iran, Turkey, Pakistan, India, Afghanistan, China, Nepal, Bangladesh, and Africa. Sporadic outbreaks were observed in a number of nations in Southern Europe and other parts of the world as a result of considerable trade with other foreign countries. (15,16). Poxviruses exploit genetic recombination to acquire host genes and escape immunity (17). One pathway is apoptosis regulator genesis, which is a controlled process of cellular death that occurs in response to external (extrinsic apoptosis) or internal (intrinsic) stimuli. Important for development, tissue homeostasis, and the elimination of damaged or pathogen-infected cells (18, 19). They are listed in group A of contagious disease by the World Organization for Animal Health (OIE) (20). This paper describes the genetic identification of sheep pox virus from field strains using phylogenetic analysis to target partial sequences of apoptosis-related genes and compare them to other global sequences, including vaccination strains.

Materials and Methods

Ethical Approval

The study was approved by the Committee for Research Ethics at the College of Veterinary Medicine, University of Al-Qadisiyah, Iraq.

Samples collection

Six hundred sheep in Diwaniyah, Iraq (Al Sannih, Daghara, and Al Badir) were evaluated, with one hundred and twenty-five of them suspected of having SPPV. This is the study that has been conducted. From September 2023 to January 2024. Skin lesion samples were collected aseptically, transported to the laboratory in a cool bag, and stored at -20°C until used for molecular testing.

Genomic DNA Extraction

Skin lesions were placed in a sterilized Petri plate and then cut into smaller pieces utilizing sterile scissors. Transfer to a sterile 1.5ml microcentrifuge tube, homogenize with tissue lysis buffer, and purify using a silica-based column per the manufacturer's instructions (ADDBio, South Korea). Eluted DNA was stored at -20°C until further examination.

Polymerase Chain Reaction (PCR)

The targeted genes were amplified using primers (Table 1) developed specifically for SPPV in this work. The reaction comprises of 20 µl of PCR master mix (ADDBio, South Korea), 1 µl of each primer (for apoptosis associated gene), and 2 µl of template viral DNA. The thermal conditions included one cycle of initial denaturation at 95 C for 10 minutes, followed by 30 cycles of denaturation at 95 C for 30 seconds, annealing at 60 C for 30 seconds, extension at 72 C for 1 minute, and one cycle of ultimate extension at 72 C for 5 minutes. PCR products were electrophoresed on an agarose gel with ethidium bromide staining and photographed using a gel documentation system (Syngene, Taiwan).

Table (1) Primers used for detection of Antiapoptotic gene

Ta rg et g e n e	Sequence '5-----3'		Am plic on size	NC BI acce ssio n nu mbe r	S ta rt	E n d
Po x- ap op to- F	Fo rw ard	GCTCGTTTAG TGCTAAATCA TCATC	253 bp	MN 072 629	1	1
	Re ver se	ATACGCGAA TGCTGTGAG GT			7	9
					4	8
					4	4
					2	0
					6	7

DNA Sequencing Method

Macrogen (South Korea) selected ten samples from each gene from the positive PCR samples for DNA Sanger sequencing. They were slightly trimmed from noise signals, phylogenetically examined, and compared to other strains from around the world. These sequences were submitted to NCBI to gain accession numbers (see Table 2). Phylogenetic analysis A phylogenetic tree and several sequence alignments were created using partial sequences from the gene of local SPPV isolates (Mega X program).

Statistical Analysis

Chi-square (X²) was used to identify substantial variations in sickness prevalence data and the impact of other factors. Variations were deemed statistically significant (P < 0.05) (21).

Results

The Endpoint PCR results were detected in 40 samples. 125 sheep of both sexes and ages were selected. The results demonstrated this (32%). Whereas 85 samples (68%) proved negative for sheepox viral DNA (Table 2). The statistical test showed a significant difference among positive and negative cases (P<0.01).

Table 2: Percentages of positive sheep pox cases by PCR

Number of sheep with skin lesions	Number of sheep	Percentage
Infected sheep	40	32
Non-Infected sheep	85	68
Total	125	100
Chi-square value	-----	32.4
P- value	-----	<0.0001(HS)

HS: Highly significant difference at P<0.01

Detection of Sheep pox genes by Conventional PCR Assay

The PCR results were detectable in 40 of the 125 suspicious sheep. Ten samples having positive PCR results were for apoptosis-related genes. After electrophoresis, the results Out of 125 skin lesions, 40 samples generated bands with expected diameters of 253 bp (Fig. 1), which corresponded to the universal ladder (50-1500 bp). Whereas 85 skin lesion samples (68%) provided negative for sheep pox virus DNA using PCR (Table 2). The statistical analysis revealed a substantial difference between positive and negative cases (P < 0.01).

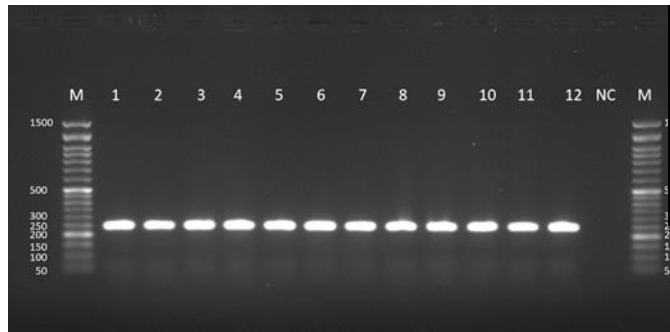


Figure (1): Agarose gel electrophoresis image (1.6 % agarose) shows positive amplicons (1-12) of sheep pox targeting partial region of antiapoptotic gene (size= 253 bp). NC is negative control in which similar PCR conditions were used except the H2O was added instead of DNA. M is molecular marker from Genedirx (South Korea).

Gene sequence and phylogenetic analysis

Ten local isolates with antiapoptotic gene sequences were deposited in the NCBI database: (OR535203), (OR535204), (OR535202), (OR535201), (OR535200), (OR535199), (OR535198), (OR535197), (OR535196), and (OR535195). These were evaluated and compared to NCBI Gen Bank sequences, which revealed some genetic differences between the detected strains and those obtained from NCBI, Table (3).

Table (3) Local SPPV isolates with their accession numbers

N o.	Obtain ed accessio n numbe r	NCBI-BLAST Homology Sequence identity (%)			
		Identic al to	Genban k Accessi on number	Count ry	Identi ty (%)
1	OR535195	Sheepp ox	MN072631	Turke y	100
2	OR535196	Sheepp ox	MN072630	Saudi Arabia	100
3	OR535197	Sheepp ox	MN072629	Canda	100
4	OR535198	Sheepp ox	MN072628	Nigeri a	100
5	OR535199	Sheepp ox	MG000157	India	100
6	OR535200	Sheepp ox	MN072626	Abu Ghari b Iraq	100
7	OR535201	Sheepp ox	MN072630	Saudi Arabia	100

3	OR535202	Sheepp ox	MN072629	Canda	100
4	OR535203	Sheepp ox	MN072628	Nigeri a	100
0	OR535204	Sheepp ox	MG000157	India	100

Phylogenetic tree and homology sequence of sheep pox virus antiapoptotic gene demonstrated a 100% agreement between the locally identified strain and the global stain (Fig. 4). ten sequences (accession no. (OR535203), (OR535204), (OR535202), (OR535201), (OR535200), (OR535199), (OR535198), (OR535197), and (OR535196) had the same identity 100% in comparison with homologues global sequence from Turkey. Table 3: Saudi Arabia, Canda, Nigeria, India, Abu Gharib _ Iraq, Nigeria, and India

Multiple sequence alignment

There are also visible conserved motives between all ten sequences motive in nucleated no. (25, 26, 27,28) - The motive in the region between nucleated NO. 56-61.



Figure (2): Multiple sequence alignment of the sheeppox virus within partial region of antiapoptotic gene. This shows similarity and differences carried out by MegaX.

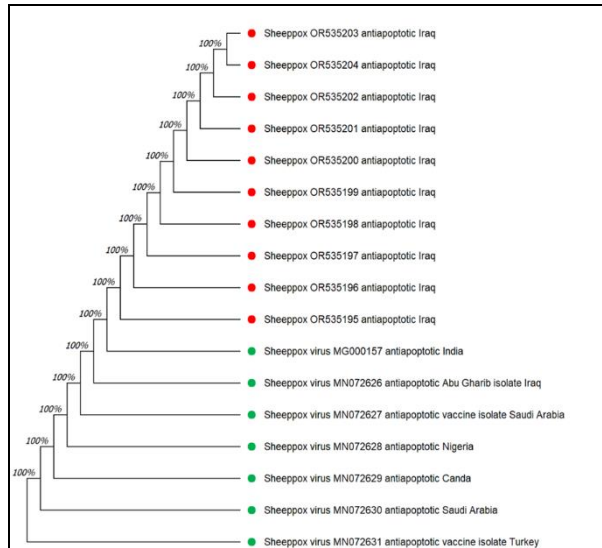


Figure (3): Evolutionary analysis by Maximum Likelihood method of sheeppox virus targeting partial region within the antiapoptotic gene.

Discussion

Capripoxvirus was isolated for the very first time in Kurdistan, northern Iraq, and identified as the GPV Sersenik strain (22). The collected DNA from skin lesions was subjected to PCR using primers specialized for antiapoptotic gene, yielding an expected amplicon of size 253 bp. There was not any amplifier in negative situations. This suggests that the primers were highly specific. To identify the sheeppox virus.

The percentage of positive sheep cases in different areas of Al-Diwaniya reached 32%. This results are consistent with other investigations, such (12) That resulted in 31.3%. Similar findings were observed by (23), who documented a morbidity rate of 36% in different ages from AL-Diwaniyah (24) In Duhok Provinces, the morbidity rate reached 30% in lambs aged 2-4 months. The infection percentage agreement could be caused by There are multiple causes such as the sickness being endemic in this location, the environment, overpopulation, management system, immune status of herds, and vaccination programmes. The conclusion was supported by offering a description of (23, 24).

In contrast, the pox viruses possess several immune escape mechanisms and encode a wide range of immune-modulating proteins (25). This contains antiapoptotic gene, as a highly effective technique to limit infections (26,27,28). This was another motive to carry out this molecular method. to see if these genes are permanent or changeable. The study employed BLAST and BioEdit software to look at the gene for similarities to the indicated strain from GenBank. The results showed a high degree of

similarity, indicating that the sequences were identical thus offering an additional Lots of confidence.

Previous studies have dealt with molecular examinations of an inhibitor of apoptosis of this gene, including a study in freshwater pearl mussel, *Hyriopsis schlegelii* in China (29), as well as the Sheep Ovary During the Reproductive Cycle, which may be involved in controlling the estrus cycle in sheep (30).

Poxviruses and herpesviruses encode secreted copies of cytokine receptors as a unique strategy for evading the host's immune response (31).

Conclusion

To the best of our knowledge, this is first study targeting apoptosis evasive gene in Iraq. It was showed that identified sequence highly similar to that in Alhaliya and Al-Ilmiyyah strains which means this gene is highly conserved. Thus, could be useful as a diagnostic gene for viral diagnosis or for vaccine development.

Acknowledgements

The authors are grateful to everyone who helped them conduct this research.

Funding source

This research had no specific fund; however, it was self-funded by the authors

Conflict of interest

No conflict of interest is disclosed by the authors.

References

- Albaroodi, S. (2018). (Isolation and pathogenesis of sheep pox virus in Nineveh governorate. December. <https://www.researchgate.net/publication/329416164>
- Sonowal, J., Lal Patel, C., Kumar Gandham, R., Sajjanar, B., Ishaq Nabi Khan, R., Ranjan Prahara, M., Akram Malla, W., Kumar, D., Dev, K., Barkathullah, N., Bharali, K., Dubey, A., Lalita, D., Zafir, I., Mishra, B. P., & Mishra, B. (2021). Genome-wide expression analysis reveal host genes involved in immediate-early infections of different sheeppox virus strains. *Gene*, 801(July), 145850. <https://doi.org/10.1016/j.gene.2021.145850>
- Sprygin, A., Shalina, K., Van Schalkwyk, A., Mazloum, A., Shcherbinin, S., Krotova, A., & Chvala, I. (2023). Molecular and Epidemiological Analyses of Sheeppox Outbreaks in Russia from 2013 to 2021. *Transboundary and Emerging Diseases*, 2023. <https://doi.org/10.1155/2023/8934280>
- Gelaye, E., & Lamien, C. E. (2019). Sheep and goat pox. *Transboundary Animal Diseases in Sahelian Africa and Connected Regions*, 289–303. https://doi.org/10.1007/978-3-030-25385-1_14



5. Zro, K., Zakham, F., Melloul, M., El Fahime, E., & Ennaji, M. M. (2014). A sheeppox outbreak in Morocco: isolation and identification of virus responsible for the new clinical form of disease. *BMC Veterinary Research*, 10(1), 1-8. <https://doi.org/10.1186/1746-6148-10-31>
6. Seyoum, B., & Teshome, E. (2017). Major transboundary disease of ruminants and their economic effect in Ethiopia. *Global Journal of Medical Research: G, Veterinary Science and Veterinary Medicine*, 17, 27-36. <https://www.researchgate.net/publication/335202253>.
7. Hurisa, T. T., Jing, Z., Jia, H., Chen, G., & He, X. B. (2018). A review on Sheeppox and Goatpox: Insight of epidemiology, diagnosis, treatment and control measures in Ethiopia. *J. Infect. Dis. Epidemiol*, 4(3), 2474-3658. DOI:10.23937/2474-3658/1510057
8. McInnes, C. J., Damon, I. K., Smith, G. L., Mcfadden, G., Isaacs, S. N., Roper, R. L., Evans, H., Damaso, C. R., Carulei, O., Wise, L. M., & Lefkowitz, E. J. (2023). ICTV Virus Taxonomy Profile: Poxviridae 2023. 1-2. <https://doi.org/10.1099/jgv.0.001849>
9. Taylor, J. M., & Barry, M. (2006). Near death experiences: Poxvirus regulation of apoptotic death. *Virology*, 344(1), 139-150. <https://doi.org/10.1016/j.virol.2005.09.032>
10. Al-Shabebi, A., El-Sabagh, I., Abu-Elzein, E., Zaghawa, A. A., Al-Naem, A. A., & Housawi, F. M. (2014). Molecular detection and phylogenetic analysis of sheeppox virus in Al-Hassa of Eastern Province of Saudi Arabia. *Adv. Anim. Vet. Sci*, 2(2S), 31-34. <http://dx.doi.org/10.14737/journal.aavs/2014/2.2s.31>
11. Constable, P. D., Hinchcliff, K. W., Done, S. H., & Gruenberg, W. (2017). A textbook of the diseases of cattle, horses, sheep, pigs, and goats. Saunders Elsevier, New York. 11th edi. P, 2217-2219. chrome-extension://efaidnbmnnnibpcajpcglclefindmkaj/http://sutlib2.sut.ac.th/sut_contents/H111504.pdf
12. Chala, A. A. (2017). Isolation and characterization of pox virus circulating in sheep and goat from outbreak cases of Aeda Berga district, West Shoa zone, central Ethiopia (Doctoral dissertation, MSc Thesis, College of Veterinary Medicine and Agriculture, Ethiopia). <https://cgspace.cgiar.org/bitstream/handle/10568/90469/Assefa.pdf?sequence=1>
13. Abd-Elfatah, E.B., El-Mekkawi, M.F., Bastawecy, I.M. and Fawzi, E.M. (2018): Identification and phylogentic analysis of sheep pox during an outbreak of sheep in Sharkia Governorate, Egypt. *Genetics and Molecular Research*, 17(2)1-12. <http://dx.doi.org/10.4238/gmr16039901>
14. Abriham, K., Etenesh, H.M. and Jiregna, D.(2018): Prevalence of Common Skin Diseases of Small Ruminants in Dibate District Metekel Zone of Benishangul Gumuz Regional State, Northwestern Ethiopia. *Multidisciplinary*. <https://www.researchgate.net/publication/324587733>
15. Constable, P. D., Hinchcliff, K. W., Done, S. H., & Grünberg, W. (2016). *Veterinary medicine: a textbook of the diseases of cattle, horses, sheep, pigs and goats*. Elsevier Health Sciences. <https://www.cabidigitallibrary.org/doi/full/10.5555/20163217871>
16. Limon, G., Gamawa, A. A., Ahmed, A. I., Lyons, N. A., and Beard, P. M. (2020). Epidemiological characteristics and economic impact of lumpy skin disease, sheeppox and goatpox among subsistence farmers in northeast Nigeria. *Frontiers in veterinary science*, 7, 8. <https://doi.org/10.3389/fvets.2020.00008>
17. Odom, M. R., Hendrickson, R. C., & Lefkowitz, E. J. (2009). Poxvirus protein evolution : Family wide assessment of possible horizontal gene transfer events. 144, 233-249. <https://doi.org/10.1016/j.virusres.2009.05.006>.
18. Taylor, J. M., & Barry, M. (2006). Near death experiences: poxvirus regulation of apoptotic death. *Virology*, 344(1), 139-150. <https://doi.org/10.1016/j.virol.2005.09.032>
19. Youle, R. J., & Strasser, A. (2008). The BCL-2 protein family: opposing activities that mediate cell death. *Nature reviews Molecular cell biology*, 9(1), 47-59. <https://doi.org/10.1038/nrm2308>
20. Rashid, P. M. A., Sheikh, M. B., Raheem, Z. H., & Marouf, A. S. (2017). Molecular characterization of lumpy skin disease virus and sheep pox virus based on P32 gene. *Bulgarian Journal of Veterinary Medicine*, 20(2), 131-40. <https://doi.org/10.15547/bjvm.984>.
21. Ab Rahman, J. (2015). Brief guidelines for methods and statistics in medical research. Springer Singapore. <https://link.springer.com/book/10.1007/978-981-287-925-7>.
22. Tantawi, H. & M. Al Falluji, 1979. Laboratory characteristics of four strains of goatpox virus. *Acta Virologica*, 23, 455-460. <https://europepmc.org/article/med/94766>
23. Ha, S., & Iq, H. (2017). Phylogenetic analysis of sheeppox virus isolates based on P32 gene in Iraq. 5(6), 704-708. www.entomoljournal.com



24. Zangana, I. K., & Abdullah, M. A. (2013). Epidemiological, clinical and histopathological studies of lamb and kid pox in Duhok, Iraq. *Bulgarian Journal of Veterinary Medicine*, 16(2), 133–138. <https://www.researchgate.net/publication/286180139>
25. Johnston, J. B., & McFadden, G. (2003). Poxvirus immunomodulatory strategies: current perspectives. *Journal of virology*, 77(11), 6093-6100
DOI: <https://doi.org/10.1128/jvi.77.11.6093-6100.2003>.
26. Upton JW, Chan FK (2014) Staying alive: cell death in antiviral immunity. *Mol Cell* 54:273–280
Upton JW, Kaiser WJ, Mocarski ES (2010) Virus inhibition of RIP3-dependent necrosis. *Cell Host Microbe* 7:302–313
<http://dx.doi.org/10.1016/j.molcel.2014.01.027>
27. Mocarski ES, Guo H, Kaiser WJ (2015) Necroptosis: the Trojan horse in cell autonomous antiviral host defense. *Virology* 479–480:160–166
Morgan MJ, Kim YS (2015) The serine threonine kinase RIP3: lost and found. *BMB Rep*48:303–312
<https://doi.org/10.1016/j.virol.2015.03.016>
28. Veyer DL, Carrara G, Maluquer de Motes C, Smith GL (2017) Vaccinia virus evasion of regulated cell death. *Immunol Lett* 186:68–80
Wallach D, Kang TB, Dillon CP, Green DR (2016) Programmed necrosis in inflammation: toward identification of the effector molecules. *Science* 352, aaf2154.
<https://doi.org/10.1016/j.imlet.2017.03.015>
29. Wu, D., Wang, C., Zhang, W., Peng, K., Sheng, J., Wang, J., ... & Hong, Y. (2019). Molecular characterization of an inhibitor of apoptosis protein (IAPs) in freshwater pearl mussel, *Hyriopsis schlegelii*. *Bioengineered*, 10(1), 365-373.
<https://doi.org/10.1080/21655979.2019.1653738>
30. Lu, Z., Tang, M., Zhang, M., Li, Y., Shi, F., Zhan, F., ... & Qin, Z. (2021). Expression and functional analysis of the BCL2-Associated agonist of cell death (BAD) gene in grass carp (*Ctenopharyngodon idella*) during bacterial infection. *Developmental & Comparative Immunology*, 123, 104160.
<https://doi.org/10.1016/j.dci.2021.104160>
31. Hernaez, B., & Alcamí, A. (2020). Virus-encoded cytokine and chemokine decoy receptors. *Current opinion in immunology*, 66, 50-56.
<https://doi.org/10.1016/j.coi.2020.04.008>.