



Sequencing strain *Fasciola* species in Cattle in Iraq

Article Info.

Author

Ismael WaadUllah Ismael

Department of Anesthesia Techniques, College of Health and Medical Techniques/ Baghdad, Middle Technical University, Baghdad, Iraq.

Corresponding Author Email Address:
ismaana.84@mtu.edu.iq

Article History

Received: Oct. 1, 2025

Accepted: Feb. 2, 2026

e Published: March 31, 2026

Article type: Research Article

<https://doi.org/10.23975/bjvr.2026.165734.1250>

Abstract

Fascioliasis is a significant zoonotic disease caused by *Fasciola* species that affects humans and animals like sheep, cows and buffaloes worldwide, causing public and veterinary health problems. The aim of this study is to focus on the genetic differentiation between the two species of *Fasciola* (*F. hepatica* and *F. gigantica*) by using the large subunit of 28S rDNA gene. A total of 45 adult worms were isolated from the liver of different cattle in Iraq-Basrah Province. Extraction of genomic DNA using a special kit was performed to isolate *Fasciola* species with 21 bp specific primers included that target 28S rDNA. The BLAST and PCR products showed evolutionary phylogenetic and genetic variation with similarity in about 80% between the two species of *Fasciola*. The results phylogenetically concluded a highly species-specific conservation and polymorphism with possible strain hybridization.

Keywords: *F. hepatica*, *F. gigantica*, Sequencing, Cattle.

Introduction

Fasciola species is a trematode parasite that has a distinct morphological appearance. The disease caused by *Fasciola* is considered an important food and water parasitic zoonosis introduced via *Fasciola* genus, which belongs to the trematode liver flukes and has a wide distribution in tropical regions that have been recorded in several countries around the world (1).

According to the public health problem of the *Fasciola* diseases, the diagnosis of the *Fasciola* species (*F. gigantica* and *F. hepatica*) is considered a primary target for determining the phenotypic and genotypic variation and application of good preventive measures for control of the disease (2).

Because of the inability to differentiate between the two species by morphological examination, a wide range of genetic approaches were developed to demonstrate the degree of differences at the DNA and RNA molecular level by using the more advanced real-time PCR, which is considered a very accurate, easy, and slightly inexpensive methods to split between *F.hepatica* and *F.gigantica* dissimilarities (3).

The second internal transcribed spacer (ITS-2) of nuclear DNA is broadly used to differentiate between the two species of *Fasciola* reported as 1.7% sequence differences (4).

Fasciolosis, according to the World Health Organisation (WHO), is considered a first parasitic disease among many infections that are distributed epidemically in many areas around the world, like Africa, Asia and Europe (5).

Fasciola life cycle starts with lying eggs which is unembryonated and excreted in the faeces of final host to the external watery environment, then these eggs transferred to the embryonated stage, in addition to hatching of miracidia inside the snail of intermediate host, inside the snail the parasite develops asexual sporocysts and cercariae, finally cercariae leave the snail in the form of mature metacercariae as a final infective stage (6,7).

The complete life cycle of *Fasciola* involves two types of hosts: the final or definitive host represented by cattle, and an intermediate host, which is a freshwater snail. The infective metacercaria can transfer to and infect humans and animals by drinking polluted water or eating contaminated watery plants containing the metacercaria (8).

In the current study, genetic heterogeneity of both *Fasciola* species isolates can be established by using a small region of the manual 28S rDNA, so the ITS region between 5.8 S and 28.5 S of a nuclear ribosomal DNA was used as a conserved sequence to differentiate the polymorphism of *Fasciola hepatica* and *Fasciola gigantica* isolates in various ITS fragments (9-11).

Materials and Methods

Parasites :45 adult worms of liver flukes were isolated from the liver and gallbladder of both slaughtered cattle in Iraq, Basrah governorate. The study was approved by the Ethical Committee of the Middle Technical University/ College of Health and Medical Techniques, Baghdad.

DNA extraction :The Qiagen/ Germany mini kit of DNA extraction (QIAamp) was used in this study for extraction of genomic DNA from the isolated flukes according to the manufacturer's instructions of the company by amplification of small subunit Gene of 28s ribosomal DNA with a molecular weight of 610bp with as well as 21 bp forward and reverse primers, finally 2.5 µl from extracted DNA was purified via a nanodrop instrument for measurement of DNA concentration.

Primer design :Primers 20 bp of F (5'-ACGTGATTACCCGCTGAACT-3') and 21 bp of R (5'-CTGAGAAAGTGCACTGACAAG-3') were used (12).

PCR :The thermocycler PCR reactions for (5 samples) were conducted with 94 ° C for 3 min, and after that 30 cycles of 30 s at 94 °C, 30 s at 56.5 °C and 60 s at 72 °C. Finally, extension of primer for 5 mins was used. A Total of 12 microliters at 72 °C. PCR products were analyzed by electrophoresis at 1.5% conc. of agarose gel. A ladder with a molecular weight of 100 bp was included and viewed by ethidium bromide.

Results

In silico validation using NCBI Primer-BLAST confirmed the specificity of the primers, producing a single amplicon of 610 base pairs that fully matched the reference sequence of *Fasciola hepatica* isolate (GenBank accession: PV082153.1). This result supports that the sequenced PCR product corresponds to the correct target gene of the intended parasite. The forward primer aligned at position 1–20 and the reverse primer at position 610–590, covering the complete amplicon region as shown in Figure (1).

```
>PV082153.1 Fasciola hepatica isolate Gymnocephalus cercaria of Ganderbal, Kashmir, 28S rDNA. large subunit ribosomal RNA gene, partial sequence

product length = 610
Forward primer 1  ACGTGATTACCCGCTGAACT  20
Template       1  ..... 20

Reverse primer 1  CTGAGAAAGTGCACTGACAAG  21
Template       610 ..... 590
```

Figure 1. In silico validation of primer pair 28F and 28R on the 28S rDNA gene of *Fasciola hepatica* (GenBank: PV082153.1). The primers produced a specific 610 bp product matching the reference sequence, confirming that the correct target gene was amplified and sequenced.

Sample code 4

Fasciola sp. isolate S10 large subunit ribosomal RNA gene, partial sequence

Sequence ID: [OR676767.1](#) Length: 597 Number of Matches: 1

Range 1: 33 to 471 [GenBank](#) [Graphics](#)

[Next Match](#) [Previous Match](#)

Score	Expect	Identities	Gaps	Strand
388 bits(429)	1e-102	352/441(80%)	2/441(0%)	Plus/Plus
Query 25	AAGTAAAAAACCCGGAATTC	CCCTTATTAACGGCCAATGAAAGGGAAAAACCCACCC	CCC	84
Sbjct 33	AAGAAACTAACCAAGGA-TTCCCTTAGTAACGGCGAGTGAACAGGGAAAAAGCCAGCACCG			91
Query 85	AAACCCGGGGTCCCTTTGGCCCTAAGCAATTTGGGGTTCAGGTTAACTCCCGAAAAATCT			144
Sbjct 92	AAACCAAGTGGTCGTTTGGCCCTAGGCAATGTGGTGTTCAGGTTAGCTCGCGAAGATGCT			151
Query 145	GCTCCCCCTAAAATCCTAAAATGGATAAGGTTACCCGGAAATGGCCCAATGAGGGTGAAA			204
Sbjct 152	GCTCCACCTTAAGTCTATAATGAGTAAGGTTACTCGGACATGGCCCAATGAGGGTGAAA			211
Query 205	GGCCCTGGGGTGGAAATTTCAAATGGCCAATATTTTCTGAACCAACCTTGGAAATCCG			264
Sbjct 212	GGCCCTGGGGTGGAGATTCAGAATGGCCAGTATCTTCTGAGCAGACCTTGGAGTCCG			271
Query 265	GTTTTTTTGAATGCAGCCCCAACGGGGTAAACTCCCTCCAAGGTAAAACTAAC			324
Sbjct 272	GTTGTTTGTGAATGCAGCCCAAAGCGGGTGGTAAACTCCATCCAAGGCTAAATACTAGCA			331
Query 325	CCAATCCAAAAAATAACCGGGGGGAAAAATTTAAAAATACTTTAAAAA			384
Sbjct 332	CGAGTCCGATAGCGAACAAAGTACCGTGAGGG-AAAAGTTGAAAAGTACTTTGAAAGAGAGAG			390
Query 385	TAaaaaaaGGGTGAAACCGTTTAAAAGTTAACAGGTGGAATTTAAATGGAAAGCTCTAAAG			444
Sbjct 391	TAAACAGTGCCTGAAACCGTTTCAAGGTAACAGGTGAGGTTGAACTGCAAGCTCTGAGG			450
Query 445	ATTTCCCTGGTAAATATGGCA	465		
Sbjct 451	ATTCAGCTGGTGAATATGGCA	471		

Sample code 19

Fasciola hepatica isolate G15 28S ribosomal RNA gene, partial sequence

Sequence ID: [HM369305.1](#) Length: 525 Number of Matches: 1

Range 1: 6 to 461 [GenBank](#) [Graphics](#)

[Next Match](#) [Previous Match](#)

Score	Expect	Identities	Gaps	Strand
456 bits(505)	2e-123	374/457(82%)	3/457(0%)	Plus/Plus
Query 31	AACGAACCCGGATTCCCTTATTAACGGCCAATGAAAGGGAAAAA	CCCCACCCCCCAACCC		90
Sbjct 6	AACTAACCAAGGATTCCCTTAGTAACGGCGAGTGAACAGGGAAAAAGCCAGCACCGAAKCC			65
Query 91	CGGGGTCTTTTGGCCCTAGGCAATGGGGGTTCAAGGTTACCTCCCGAAAAATCCTGCTCC			150
Sbjct 66	TGTGGTCATTTGGCCCTAGGCAATGTGGTGTTCAGGTTAGCTCGCGAAGATGCTGCTCC			125
Query 151	CCCCAATTCCTTAAATGATTAAAGGTTACCCGGAAATGGCCCAATGAGGGTGAAAGGCC			210
Sbjct 126	ACCTAAGTCTATAATGAGTAAGGTTACTCGGACATGGCCCAATGAGGGTGAAAGGCC			185
Query 211	CGGGGGTGGAAATTCAAAATGGCCAATTTTTTCTGAACGAACCTTGGAAATCGGGTTTT			270
Sbjct 186	GTGGGGTGGAGATTCAGAAKGGCCAGTATCTTCTGAGCARACCTTGGAGTGGGTTGT			245
Query 271	TTTTGAATGCACCCCAAACCGGGGGTAAACTCCTTCCAAGGCTAAATACTAACCCaaat			330
Sbjct 246	TTGTGAATGCASCCCAAAGCGGGTGGTAAACTCCWTCCAAGGCTAAATACTAGCACRAGT			305
Query 331	CAAAAAAATAACCGGGGGGAAAAATTTAAAAATACTTTGAAAAAAGTAAAAA			390
Sbjct 306	CCRATAGCGAACAAAGTACCGTGAGGGAAAGTTGAAAAGTACTTTGAARAGAGAGTAAACA			365
Query 391	TTTGTGAAACCGTTTGAAGGTAAACAGTGTGG-GTTGATTTG-AAGTTTTGAGGATTTA			448
Sbjct 366	GTGCGTGAACCGTTTCAAGGTAAACAG-GTGGAGTTGAACTGCAAGCTCTGAGGATTCA			424
Query 449	CTTGGTGAATATGGCATAAACTTGGTCATATTGGTTG	485		
Sbjct 425	SCTGGTGAATATGGCATGAGCTTGGTCATATTGGTTG	461		

Sample code 45: No significant similarity found.

Sample code 50

Fasciola gigantica gene for 28S rRNA, partial sequence, isolate: KhuzBufa2G

Sequence ID: [AB674553.1](#) Length: 580 Number of Matches: 1

Range 1: 11 to 460 [GenBank](#) [Graphics](#)

[▼ Next Match](#) [▲ Previous Match](#)

Score	Expect	Identities	Gaps	Strand
453 bits(502)	8e-123	372/450(83%)	2/450(0%)	Plus/Plus
Query 39	ACCCGGATTCCCTTAATAA	GGCCAATGAA-AGGGAAAAA	ccccccccAAACCCGGGG	96
Sbjct 11	ACCAGGATTCCCTTAGTAA	CGGCGAGTGAACAGGGAAA	AGCCAGCACCGAAGCCTGT	70
Query 97	TCCTTTGGCCCTAAGCAAT	GTGGTTCAGGTTAACTCCCG	AAAAATCTGCTCCCCCT	156
Sbjct 71	TCGTTTGGCCCTAGGCAAT	GTGGTTCAGGTTAGCTCGCG	AAGATGCTGCTCACCCCT	130
Query 157	AAATCCTATAATGAATAAG	GTTACCCCGAAATGGCCCA	ATGAGGGTGAAAGGCCCT	216
Sbjct 131	AAGTCTATAATGAGTAAG	GTTACTCGGACATGGCCCA	ATGAGGGTGAAAGGCCCT	190
Query 217	GGTGGAAATTTCAAAATGG	CCAATATCTTCTTGAACGG	ACCTTGGAATCCGGttttt	276
Sbjct 191	GGTGGAGATTCAGAAATGG	CCAATATCTTCTTGAACGG	ACCTTGGAATCCGGTGT	250
Query 277	AATGCAACCCAAAACGGGG	GGTAAACTCCCTCCAAGGG	TAAATACTAACCcaaatccc	336
Sbjct 251	AATGCAGCCAAAAGCGGG	TGGTAAACTCCATCCAAGG	CTAAATACTAGCACGAGTCC	310
Query 337	aacgaaaaataaccgtgagg	gaaaaatttaaaaatacttt	taaaaaaaaataaaaaaTTGG	396
Sbjct 311	AGCGAACAAAGTACCGTG	AGGGAAAGTTGAAAAGTACT	TTGAAAGAGAGAGTAAACAG	370
Query 397	TGGAACCGTTTAAAGGTAA	AAAAGTGGAGTTTAACTT	GAAGCTTTGAGGATTTACCT	456
Sbjct 371	TGAAACCGTTTAAAGGTAA	CAAGTGGAGTTGAACTGCA	AGCTCTGAGGATTCAGCTGG	430
Query 457	GAATATGGCATGAACCTTT	GTCATATTGGTT	486	
Sbjct 431	GAGTATGGCATGAGCTTGG	TCAATTGGTT	460	

sample code 73

Fasciola hepatica isolate G15 28S ribosomal RNA gene, partial sequence

Sequence ID: [HM369305.1](#) Length: 525 Number of Matches: 1

Range 1: 15 to 460 [GenBank](#) [Graphics](#)

[▼ Next Match](#) [▲ Previous Match](#)

Score	Expect	Identities	Gaps	Strand
409 bits(453)	3e-109	356/447(80%)	1/447(0%)	Plus/Plus
Query 31	GGATTCCCTTATTAACGGCA	AATGAAAGGGAAAAA	ccccccccAAACCCGGGGGTCT	90
Sbjct 15	GGATTCCC-TTAGTAACGGC	GAGTGAACAGGGAAA	AGCCAGCACCGAACKCTGTGGTCA	73
Query 91	TTTGGCCCTTAGGCAATTGG	GGGTTTCAGGTTACCTCCCA	AAAAATCCTGCTCCCCCTAAT	150
Sbjct 74	TTTGGCCCTTAGGCAATGT	GGTGTTCAGGTTAGCTCGCG	AAGATGCTGCTCACCCCTAAG	133
Query 151	TCCTTTAATGATTAAGGTTAC	CCGGAAATGGCCCAATGAG	GGTGAAAGGCCCGGGGGT	210
Sbjct 134	TCCTATAATGAGTAAGGTTAC	TCGGACATGGCCCAATGAG	GGTGAAAGGCCCGTGGGGT	193
Query 211	GGAAATTCAAAATGGCCAT	TTTTCTTCCATAAACAAAC	CTTGGAATCGGGttttttGAAT	270
Sbjct 194	GGAGATTCAGAAKGGCCAG	TATCTTCTGAGCARACCT	TGGAGTCGGGTGTTTGTGAAT	253
Query 271	GCACCCCAAACCGGGGGTAA	ACTCCTTCCAAGGCTAAAT	ACTAACCcaatccaaaacc	330
Sbjct 254	GCASCCCAAAGCGGGTGGT	AAACTCCWTCCAAGGCTAA	ATACTAGCACRAGTCCRATAGC	313
Query 331	aaaaaataaccgggagggaa	atttaaaaatactttgaaaa	aaaaataaaaCATTTGTTGA	390
Sbjct 314	GAAACAAGTACCGTGAGGG	AAAGTTGAAAAGTACTTTG	AARAGAGAGTAAACAGTGC	373
Query 391	AACCGTTTAAAAGTAAAAG	TTAAGTTAATTGCAAGTT	TAAAGTTTTACTTGGTAAT	450
Sbjct 374	AACCGTTTCARAGGTAAAC	AGGTGGAGTTGAACTGCA	AGCTCTGAGGATTCASCTGG	433
Query 451	TATGGCATAACCTTGGTCTA	ATTGGTT	477	
Sbjct 434	TATGGCATGAGCTTGGTCA	TATTGGTT	460	

Table (1): BLASTn identification of *Fasciola* spp. based on sequence analysis.

Sample ID	Top Match Organism	Accession No.	Identity (%)	Score (bits)	E-value
4_ISF	<i>Fasciola</i> sp. isolate S10	OR676767.1	80%	388	1e-102
19_ISF	<i>Fasciola hepatica</i> isolate G15	HM369305.1	82%	456	2e-123
50_ISF	<i>Fasciola gigantica</i> isolate KhuzBufa2G	AB674553.1	83%	453	8e-123
73_ISF	<i>Fasciola hepatica</i> isolate G15	HM369305.1	80%	409	3e-109

The obtained 28S rDNA sequences from four PCR-positive samples (4_ISF, 19_ISF, 50_ISF, and 73_ISF) were analyzed using BLASTn against the NCBI nucleotide database. The results confirmed that all sequences belonged to the genus *Fasciola*, with varying degrees of identity.

Sample 4_ISF showed 80% identity with *Fasciola* sp. isolate S10 (GenBank: OR676767.1), confirming the genus-level classification but with possible sequence variability or subspecies representation.

Sample 19_ISF showed 82% identity with *Fasciola hepatica* isolate G15 (GenBank: HM369305.1) with a score of 456 bits and an E-value of 2e-123. Similarly, sample 73_ISF matched *Fasciola hepatica* (HM369305.1) with 80% identity, while sample 50_ISF aligned most closely with *Fasciola gigantica* (GenBank: AB674553.1) at 83% identity, suggesting species-level differentiation.

These results demonstrate the existence of both *F. hepatica* and *F. gigantica* lineages in the examined samples and validate the specificity of the 28S rDNA target region for molecular detection of *Fasciola* species.

Four *Fasciola* samples (4_ISF, 19_ISF, 50_ISF, and 73_ISF) were compared to the reference sequence *Fasciola hepatica* isolate *Gymnocephalus cercaria* (GenBank: PV082153.1) using multiple sequence alignment of the 28S rDNA region. High levels of sequence conservation were found throughout most regions in the alignment, particularly in samples 19_ISF and 73_ISF. Samples 4_ISF and 50_ISF, on the other hand, showed several nucleotide changes, suggesting species-level variation. The identification of *F. hepatica*, *F. gigantica*, and *Fasciola* sp. among the examined isolates is supported by these mutations, which are in line with earlier BLAST results. (Fig. 2)

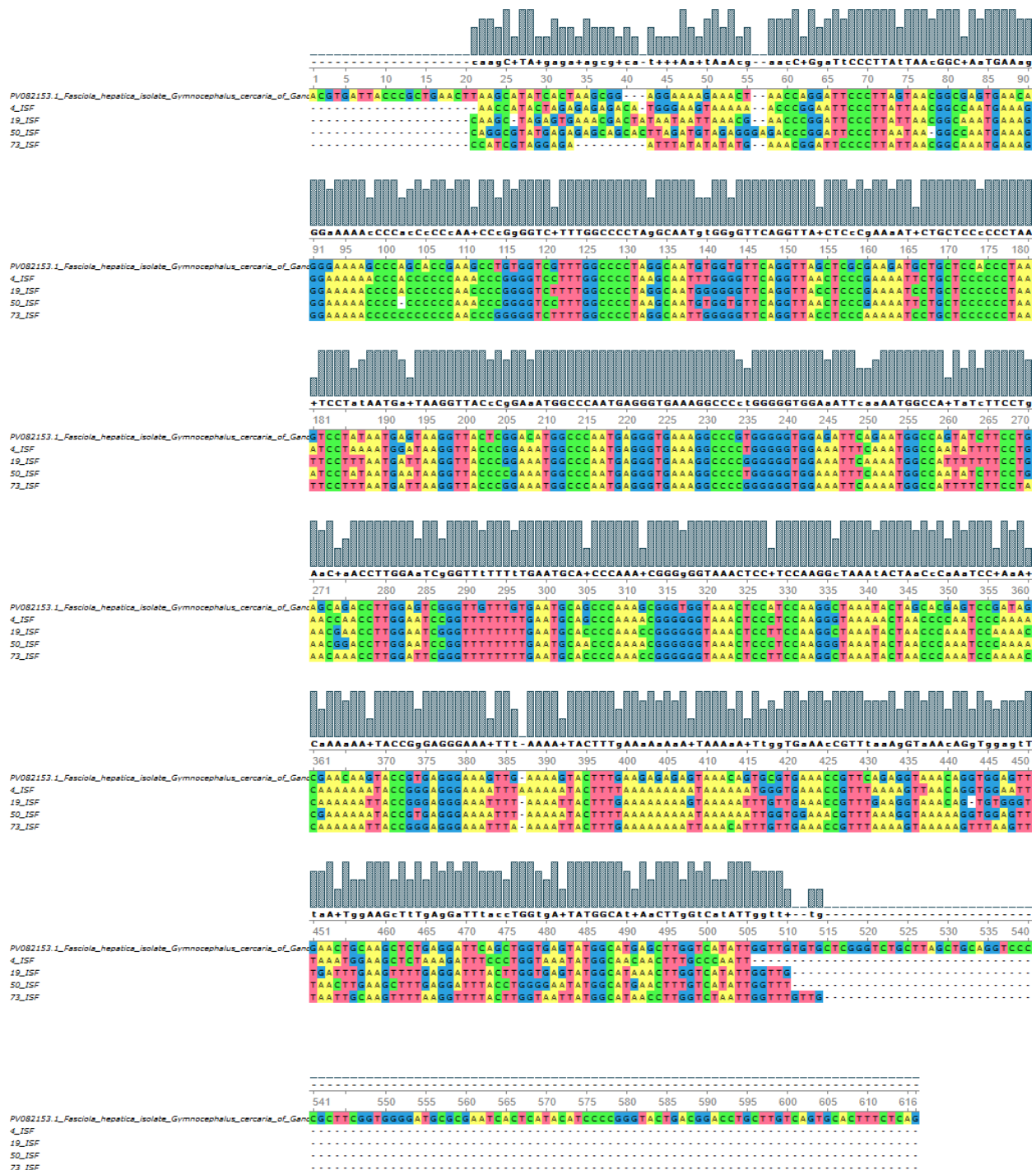


Figure 2. 28S rDNA sequences from four *Fasciola* isolates (45_ISF, 19_ISF, 50_ISF, and 73_ISF) were aligned many times and compared to the reference *Fasciola hepatica* isolate (PV082153.1). Sequence conservation or variation is indicated by colored nucleotides. High conservation is reported in most regions, whereas substitutions are detected in certain places, particularly in samples 45_ISF and 73_ISF.



Figure 4. Multiple sequence alignment of translated 28S rDNA amino acid sequences from four *Fasciola* isolates and the reference sequence *Fasciola hepatica* (PV082153.1). Conserved regions are indicated by identical amino acids across samples, while substitutions highlight possible functional or evolutionary differences.

Discussion

Genetic specialization of *Fasciola* species based on a large subunit gene of 28S ribosomal DNA provides significant analysis in terms of genetic variability of Iraqi *Fasciola* species. These study results provide a significant specificity of primers that trigger the targeted genes, giving more precise results along with ribosomal DNA of the parasite's fluke (12,13). In the Middle East regions, the BLAST analysis showed different identity levels about *F. hepatica* isolates, whereas it showed very high similarity about *F. gigantica*, these results interpret a group distribution of the 2 species in the epidemic regions (14,15). In previous genetic studies, the nucleotide substitution level of 45 ISF and 73 ISF was reported for intraspecific hybridisation and variability (16,17). The presence of a very high conservation degree of 28s ribosomal DNA gene, mainly between *Fasciola hepatica* isolates, as well as polymorphism was significant in certain gene sequences reporting intraspecies variation level, there are a cluster of isolates with phylogenetic analysis about *Fasciola hepatica* of a reference gene, whereas the presence of a few co-existence of a distinct divergent genetically lineages (14,16). The sequence of amino acid analysis showed a highly conserved domain of protein constructing a physicochemically conserved ribosomal subunit, in addition to a small level of genetic substitution in some isolates, reporting mechanisms of adaptation to evolution that affect a parasite's habit, host shelter specificity and treatment sensitivity (18,19). Finally, our results regarding genetic variation and intraspecific complexity of *Fasciola* studied species revealed the essential surveillance and investigations at the genetic level, focusing on the hybridization monitoring and species differentiation. Effective measurements to parasitic infection are considered a good strategy to control epidemiology, pathogenicity and outcome of treatments (17,20).

Conclusions

The molecular results of this study targeting 28S rDNA of *Fasciola* species isolates revealed the genetic variation and coexistence of the two species with genetically evolutionary divergent strains. The primers of this study were very species-specific and efficient for targeting the amplification of this gene. The phylogenetic classification analysis showed highly conserved polymorphism with variation in strain level hybridization.

Acknowledgment

The authors are thankful to the College of Health and Medical Techniques- Research Unit in Baghdad, Iraq.

Conflict of Interests

The authors declare that they have no potential conflict of interest with respect to the publication of this article.

Ethical Clearance

This work is approved by The Research Ethical Committee.

References

1. Ai, L., Chen, M. X., Alasaad, S., Elsheikha, H. M., Li, J., Li, H. L., ... & Chen, J. X. (2011). Genetic characterization, species differentiation and detection of *Fasciola* spp. by molecular approaches. *Parasites & vectors*, 4(1), 101. <https://doi.org/10.1186/1756-3305-4-101>.
2. Sedighe, M. I. R., Dabirzadeh, M., Rokni, M. B., Aryaeipour, M., Shahraki, M. K., & Azizi, H. (2019). Identification and phylogenetic classification of *Fasciola* species isolated from sheep and cattle by PCR-RFLP in Zabol, in Sistan and Baluchistan province, Southeast Iran. *Iranian Journal of Public Health*, 48(5), 934.
3. Heydarian, P., Jajarmi, V., Spotin, A., Ashrafi, K., Mohebbi, M., Aryaeipour, M., ... & Rokni, M. B. (2022). Molecular characterization of animal *Fasciola* spp. isolates from Lorestan Province, Western Iran. *Iranian Journal of Public Health*, 51(8), 1847. doi: [10.18502/ijph.v51i8.10271](https://doi.org/10.18502/ijph.v51i8.10271)
4. Khalafala, R. E. (2020). Prevalence and phylogenetic analysis of *Fasciola* species in upper Egypt based on ribosomal ITS-2 gene sequencing. *Egyptian Veterinary Medical Society of Parasitology Journal (EVMSPJ)*, 16(1), 142-158 DOI :[10.21608/EVMSPJ.2020.132162](https://doi.org/10.21608/EVMSPJ.2020.132162)
5. Omar, M. A., Elmajdoub, L. O., Ali, A. O., Ibrahim, D. A., Sorour, S. S., Al-Wabel, M. A., ... & Metwally, A. M. (2021). Genetic characterization and phylogenetic analysis of *Fasciola* species based on ITS2 gene sequence, with first molecular evidence of intermediate *Fasciola* from water buffaloes in Aswan, Egypt. *Annals of Parasitology*, 67(1). doi: [10.17420/ap6701.312](https://doi.org/10.17420/ap6701.312)
6. Koyee, Q. M., Khailany, R. A., Rahman, M. L., & Nassraddin, L. N. (2024). Histopathologic changes and molecular characterization of fascioliasis (a zoonotic disease) among slaughtered livestock in Erbil and Halabja Abattoirs, Kurdistan region-Iraq. *Baghdad Science Journal*, 21(7), 2191-2191. <https://doi.org/10.21123/bsj.2023.9099>
7. Raouf, H. S., Marif, H. F., Rahman, H. S., Omar, M., Sheikh, B., & San Ahmed, A. M. (2020). Molecular characterization and phylogenetic analysis of *Fasciola* species in sheep and goats in Sulaymaniyah Province, Northern Iraq. *JZ S*, 22(1), 297-305. doi: [10.17656/jzs.10794](https://doi.org/10.17656/jzs.10794)
8. Rekani, A. M., & Mero, W. M. (2023). Molecular characterization of *Fasciola* spp. from ruminants in Duhok province using the ITS1 ribosomal DNA marker. *Iraqi J Vet Sci*. 37(2):315-23. doi: [10.33899/IJVS.2022.134290.2358](https://doi.org/10.33899/IJVS.2022.134290.2358).
9. Othman, V. S., & Hama, A. A. (2023). Prevalence and molecular characterization of liver fluke in Sulaimani Province, Kurdistan, Iraq. *Journal of Entomological Research*, 47(3), 615-620. DOI: [10.5958/0974-4576.2023.00113.5](https://doi.org/10.5958/0974-4576.2023.00113.5)

10. Hamoo, R. N., Al-Rubaye, F. S. I., & Mustafa, N. G. (2019). Molecular characterization and phylogenetic analysis of *Fasciola gigantica* in Iraqi sheep using ITS1. *Adv. Anim. Vet. Sci*, 7(4), 256-260. <http://dx.doi.org/10.17582/journal.aavs/2019/7.4.256.260>
11. Essa, I. M., Azzal, G. Y., & Abdulwahid, A. T. (2024). Molecular sequencing analysis of fasciola spp. in sheep. *Adv. Anim. Vet. Sci*, 12(10), 1846-1852. <https://dx.doi.org/10.17582/journal.aavs/2024/12.10.1846.1852>
12. Othman, V. S., Hama, A. A., Zorab, R. H., & Dalimi, A. (2023). Molecular Characterization of Liver Fluke Isolated from Sheep, Goat and Cattle in Sulaymaniyah, Iraq. *Iranian Journal of Parasitology*, 18(4), 554. doi: [10.18502/ijpa.v18i4.14264](https://doi.org/10.18502/ijpa.v18i4.14264).
13. Zeng, M., Wang, X., Lan, Z., Guo, X., Jiang, Y., Wu, T., ... & Wang, C. (2022). Identification of new polymorphic positions in rDNA sequences of the “intermediate” *Fasciola* forms. *Parasitology international*, 88, 102555. <https://doi.org/10.1016/j.parint.2022.102555>
14. Nukeri, S., Malatji, M. P., Sengupta, M. E., Vennervald, B. J., Stensgaard, A. S., Chaisi, M. E., & Mukaratirwa, S. (2022). Potential hybridization of *Fasciola hepatica* and *F. gigantica* in Africa. *Pathogens*, 11(11), 1303 [https://DOI.org/10.3390/pathogens11111303](https://doi.org/10.3390/pathogens11111303).
15. Hansh, W. J. (2024). Morphological and phylogenetic characterization of fasciola species isolated from cows and buffaloes in Thi-Qar province. *Iraq. J. Anim. Health Prod*, 12(1), 40-47. <http://dx.doi.org/10.17582/journal.jahp/2024/12.1.40.47>
16. Modabbernia, G., Meshgi, B., & Kinsley, A. C. (2024). Climatic variations and *Fasciola*: a review of impacts across the parasite life cycle. *Parasitology Research*, 123(8), 300. <https://doi.org/10.1007/s00436-024-08319-6>.
17. Yihunie, D. T., Mugisha, J. Y., Gebru, D. M., & Alemneh, H. T. (2024). Optimal control and cost-effectiveness analysis of *Fasciola hepatica* model. *Heliyon*, 10(19). doi: [10.1016/j.heliyon.2024.e38540](https://doi.org/10.1016/j.heliyon.2024.e38540) [External Link](#)
18. Flores-Velázquez, L. M., Ruiz-Campillo, M. T., Herrera-Torres, G., Martínez-Moreno, Á., Martínez-Moreno, F. J., Zafra, R., ... & Pérez, J. (2023). Fasciolosis: pathogenesis, host-parasite interactions, and implication in vaccine development. *Frontiers in veterinary science*, 10, 1270064. <https://doi.org/10.3389/fvets.2023.1270064>
19. Cwiklinski, K., & Dalton, J. P. (2018). Advances in *Fasciola hepatica* research using ‘omics’ technologies. *International journal for parasitology*, 48(5), 321-331. <https://doi.org/10.1016/j.ijpara.2017.12.001>
20. Chiappetta, V., Cantón, C., Pruzzo, C., Lanusse, C., Alvarez, L., & Ceballos, L. (2025). Albendazole and Clorsulon in *Fasciola hepatica* Control: Integrated Pharmacokinetic and Flukicidal Efficacy Assessment in Sheep. *Journal of Veterinary Pharmacology and Therapeutics*, 48(6), 474-483. <https://doi.org/10.1111/jvp.70009> [Digital Object Identifier \(DOI\)](#)

تتابع تسلسل جين الحمض النووي الريبوزي (rDNA) لجنس المتورقة في الماشية في العراق

إسماعيل وعد الله إسماعيل

قسم تقنيات التخدير، كلية التقنيات الصحية والطبية/ بغداد، الجامعة التقنية الوسطى، بغداد، العراق.

الخلاصة

داء المتورقة مرض حيواني المنشأ خطير، تسببه أنواع المتورقة التي تصيب الإنسان والحيوانات مثل الأغنام والأبقار والجاموس في جميع أنحاء العالم، مما يسبب مشاكل صحية وبيطرية عامة. ركزت هذه الدراسة على التمايز الجيني بين نوعي المتورقة (*F. gigantica*, *F. hepatica*) باستخدام وحدة فرعية كبيرة من جين (28S rDNA) تم عزل 45 دودة بالغة من أكباد الأبقار مختلفة في محافظة البصرة العراقية، وتم استخراج الحمض النووي الجيني باستخدام مجموعة خاصة على أنواع المتورقة المعزولة مع تضمين بادئات محددة بطول 21 زوج قاعدي تستهدف (28S rDNA) ظهرت نتائج PCR و BLAST تطورا وتنوعا وراثيا مع تشابه بنسبة 80% تقريبا بين نوعي المتورقة. خلصت النتائج من الناحية التطورية إلى حفظ وتعدد أشكال عالية الخصوصية للأنواع مع احتمال تهجين السلالات.

الكلمات المفتاحية: *F. hepatica*، *gigantica*، تتابع، الماشية.